



Effects of *Aspergillus flavus* inoculation on spring Maize (*Zea mays* L.) grain yield

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Abstract

Maize (*Zea mays* L.) is a staple crop and widely consumed over the globe, but its production is very challenging due to its contamination by *Aspergillus* species. *Aspergillus flavus* is the most dominant fungi that produce aflatoxin in many major crops. Aflatoxin producing fungi mostly attack during the development of crops and induce infection. Present study was aimed to investigate the effects of *A. flavus* inoculation on grain yield of maize genotypes in open field environments at pre-harvest time. Fourteen maize genotypes were treated with *A. flavus* inoculum by non-wounding method (inoculated through cob silk), at grain filling stage and yield related parameters were recorded. Results indicated that maize genotype R-3305 produced higher values for cob length and number of grains per cob. Maize genotype FH-949 produced higher values for cob diameter and cob weight. Maize genotype FH-1046 produced higher value for total grains weight per cob and KSC-9663 produced higher values for 100 grains weight as compared to all other genotypes. HC-2040 produced lower values for yield related parameters as compared to all other genotypes. Based on the results of current work, it can be concluded that *A. flavus* inoculation affected the grain yield related parameters. The maize genotype, FH-949 and FH-1046 produced better grain yield as compared to all other genotypes. Thus *A. flavus* inoculation affected the grain yield related component and had a pronounced role in the reduction of maize production.

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Introduction

Maize having a major role in economy of numerous countries (Khan *et al.*, 2015) is consumed as food for human and feed for livestock as well as poultry (Shakoor *et al.*, 2017). Maize is important source of proteins, carbohydrates, minerals and antioxidants which are essential components of daily nutrition and due to these qualities it has got great attention (Kuhn *et al.*, 2010; Hiran *et al.*, 2016; Ogbaga *et al.*, 2017). Focusing the importance of maize in economic, food, feed and biofuel, there is intensive requirement to understand that how crop tolerates stress and reduce its effects that cause reduction in crop production (Ogbaga *et al.*, 2017).

Aspergillus genus is ubiquitous and is found in air, soil and in debris (Maina *et al.*, 2016). Numerous factors like moisture, drought stress, high temperature and delayed harvesting favour the infection of *Aspergillus* species and aflatoxin contamination (Atanda *et al.*, 2011; Maina *et al.*, 2016). Aflatoxin producing fungi infected the maize at different stages including pre-harvest, harvest and post-harvest stage and disturb the country's economy (Atanda *et al.*, 2011), thus maize production is very challenging due to the colonization by *Aspergillus* species (Wagacha and Muthomi, 2008).

A. flavus is most dominant fungi that produce aflatoxin in many major crops. Aflatoxigenic fungi mostly attack during development of crop and caused pre-harvest aflatoxin contamination, high temperature and other stresses favour its infection and aflatoxin contamination (Farfan *et al.*, 2015).

Climate change and its impacts have become more and more prominent from past few decades (Patt and Schroter, 2008). Climate change projections propose a more changing climate with higher susceptibilities in developing countries. Pakistan is considered as the most vulnerable country with respect to climate change because it has limited resources and geographical pattern (Schilling *et al.*, 2013). Pakistan is already facing the severity of climate change events like extreme temperature, drought and floods which

ultimately resulted in high risk of infections and diseases (Smit and Skinner, 2002).

Aflatoxin contamination in the maize plant is very serious agricultural problem especially under high temperature and drought (Fountain *et al.*, 2014). Consumption of food and feed that is contaminated with aflatoxin, has been shown to affect child development, increase death rate in farm animals and liver cancer in human beings, or even cause aflatoxicosis and death in rare cases (Probst *et al.*, 2010). It was estimated that over 55 million people worldwide are exposed to aflatoxins at unhealthy levels (Liu and Wu, 2010). Due to these health risks, several countries have developed the strict rules regarding the permissible levels of aflatoxin in food and feed (Wu and Munkvold, 2008).

Plants have numerous host defense mechanisms that provide protection to the cell against stresses (Gong *et al.*, 2001; Foyer and Noctor, 2003; Pastori *et al.*, 2003; Wahid *et al.*, 2007). Plant susceptibility to aflatoxin is very complex feature and complete source of maize resistance is not known (Mayfield *et al.*, 2012; Farfan *et al.*, 2015). In addition to complexity of the colonization and aflatoxin production have strong host and pathogen interactions (Amaike and Keller, 2011; Christensen and Kolomiets, 2011; Kelley *et al.*, 2012).

Keeping in view the importance of maize, current study was designed to assess the resistance of different maize genotypes to *A. flavus* and its influence on yield characteristics of field grown maize. In this study maize genotype(s), which were the superior in term of yield with resistance against *A. flavus* contamination were assessed to investigate the effects of *A. flavus* inoculation on grain yield of maize genotypes in open field environments at pre-harvest time, in Pakistan.

Materials and methods

Field experiment was set up under semi-arid climate of Pakistan in 2017, in the field of Ayub Agricultural Research Institute (AARI), Faisalabad, Pakistan. This

study involved total fourteen maize genotypes Pearl, MMRI, HC-9091, HC-2040, YH-1898, R-2315, R-33334, R-3305, R-2207, K.S.C 9663, K.S.C 9618, FH-949, FH-1046 and Malka-16 from Pakistan. Experiment was carried out in a randomized complete block design with five replications. Each plot had size of 5.0 x 0.75 m and inter row distance was kept 20 cm while inter plant distance was 75 cm. All cultural and agronomic practices were carried out as farmer used routinely.

A. flavus were cultured on the potato dextrose agar plate for 5-7 days at 28°C and identified morphologically (Klich, 2002). The culture was prepared in broth potato dextrose and 1×10^4 conidial concentration was made, that was enough for producing aflatoxin contamination in maize grains (Windham and Williams, 2016).

The fungal inoculums were applied to the cob through silk at grain filling stage. After applying the treatment of *A. flavus*, the rows were divided into two parts; non-inoculated and inoculated with *A. flavus*. After maturation of kernels, cobs were harvested by hand and data was recorded.

Meteorological data

The meteorological data, including temperature, rainfall and relative humidity of growing time from August to December, was obtained from the climatic station (about 300m from field) of the Department of Climate Change, Ayub Agriculture Research Institute Faisalabad (Pakistan). This data is presented in the Table 1.

Yield and yield-related parameters

Cob weight

Cob weight (g) was measured using a digital weighing balance (Ashraf *et al.*, 2016).

Cob length

To measure this physical trait, a total of 5 plants were selected and their lengths (cm) were measured using a measuring tape, and then an average value was recorded (Ashraf *et al.*, 2016).

Cob diameter

Cob diameter (cm) was measured by vernier caliper (Ashraf *et al.*, 2016).

Number of grains per Cob

Number of grains per cob was counted manually (Ashraf *et al.*, 2016).

Total Grain Weight per Cob

Total grain weight per cob (g) was measured in grams using a digital weighing balance (Ashraf *et al.*, 2016).

100 Grains Weight

Cob weight (g) was measured using digital weighing balance (Ashraf *et al.*, 2016).

Statistical analysis

The data was statistically analyzed by analysis of variance using Minitab-17 software and correlation by using SPSS version 24. The significance was determined at 0.05 probability level for each treatment.

Results

Meteorological data

Table 1 shows that the climatic conditions *viz.*, air temperature (10–35 °C), relative humidity (46–87 %) and rain fall (1.1–3.8 mm) recorded in the current study were favorable for fungal growth of *A. flavus* inoculation.

Yield related parameters

Cob length

Maize genotype, R-3305 produced highest cob length while maize genotype R-2315 and HC-2040 produced the lowest cob length (Fig. 1).

The *A. flavus* inoculated and non-inoculated plants showed a differential response with respect to cob length. Analysis of variance showed that for cob length there were highly significant differences among genotypes ($P < 0.001$) and significantly differences for treatments ($P < 0.05$), whereas Genotypes x Treatments interactions showed non-significant differences ($P > 0.05$; Table 2).

Cob diameter

Maize genotype FH-949 produced the highest cob diameter followed by the maize genotype MMRI and Pearl (Fig. 2). The *A. flavus* inoculated and non-

inoculated plants showed a differential response with respect to cob diameter, which showed that cob diameter, was also not affected by the inoculation.

Table 1. Monthly average temperature, relative humidity and rain fall during the growing season of maize crop.

Months	Air Temperature (°C)		Relative Humidity (%)		Rain fall (mm)
	Max	Min	8:00 am	5:00 pm	
August	37.4	26.9	73.1	54.9	65.7
September	36.9	23.4	69.9	51.2	19
October	35.1	18.6	71.8	46.4	-
November	24.4	10.9	87.4	63.3	1.1
December	23.4	5.4	84.1	50	3.8

Analysis of variance showed that there were highly significant differences among genotypes and Genotypes x Treatments interactions ($P < 0.001$) and significantly different ($P < 0.01$) for treatments for cob diameter (Table 2).

Cob weight

Highest cob weight was observed in FH-949, followed by FH-1046, while maize genotype HC-2040 followed

by HC-9091 and KSC-9618 produced the lowest value for cob weight (Fig. 3). Results showed that inoculum had reduced the cob weight.

Analysis of variance showed that cob weight had highly significant difference for genotypes and Genotypes x Treatments interactions ($P < 0.001$) and significantly different ($P < 0.01$) for treatments (Table 2).

Table 2. Analysis of variance for yield related characteristics of fourteen different maize genotypes under treatment of *A. flavus*.

	Df	Cob L	Cob D	Cob W	No. of grains /cob	Total grains W/ cob	100 grains W
Genotypes	13	6.68***	1.75***	15859***	78929***	6102.7***	327.98***
Treatments	1	0.16	0.63**	2979**	257572***	10393.6***	860.12***
Genotypes*	13	0.77*	0.84***	2900.8***	18935***	482.2*	62.28***
Treatments							
Error	112	0.78	0.11	362.8	5865	248.1	0.95
Total	139						

L=Length; D= Diameter; W= Weight;

(*, **, *** Means square significant at $P < 0.05$, $P < 0.01$ and $P < 0.001$, respectively.).

Number of grains per cob

Maize genotype R-3305 and MMRI produced the highest value for number of grains per cob followed by R-3334 and FH-1046 (Fig. 4). While the maize genotype, HC-2040 had lowest value for the number of grains per cob. Number of grains per cob was higher in non-inoculated plants as compared to the inoculated plants. Analysis of variance showed that there were highly significant differences among

genotypes, treatments and Genotypes x Treatments interactions ($P < 0.001$) for number of grains per cob (Table 2).

Total grains weight per cob

Maize genotype FH-1046 produced the highest value for total grains weight per cob followed by FH-949 Pearl and YH-1898 (Fig. 5). Maize genotypes HC-2040 and HC-9091 produced the lowest value for the

total grains weight per cob. Overall all inoculated plants showed decrease in total grain weight per cob as compared to the non-inoculated plants except HC-9091. Analysis of variance showed that there were

highly significant differences among genotypes and treatments ($P < 0.001$) whereas Genotypes x Treatments interactions had significant differences ($P < 0.05$) for total grains weight per cob (Table 2).

Table 3. Pearson correlation between maize genotypes, treatment of *A. flavus* and yield related traits.

	Treatments	Cob's Length	Cob's diameter	Cob's Weight	No. of Grains/Cob	of Total weight/Cob	grains 100 Grains Weight
Genotypes	0.87**	0.04	-0.08	-0.13	-0.33**	-0.295**	-0.29**
Treatments		0.030	0.116	-0.102	-0.34**	-0.29**	-0.38**
Cob's Length			0.099	0.22**	0.296**	0.32**	0.04
Cob's diameter				0.29**	0.11	0.26**	0.16
Cob's Weight					0.53**	0.75**	0.54**
No. of Grains/Cob						0.75**	0.21*
Total grains weight/Cob							0.57**

(Pearson correlation* and ** is significant at 0.05 and 0.01 probability level respectively).

100 grains weight

100 grains weight was affected by the inoculation and non-inoculated plants had higher values as compared with the non-inoculated plants (Fig. 6). Maize

genotype KSC-9663 produced highest value for 100 grains weight followed by the FH-1046, Malka-16 and Pearl.

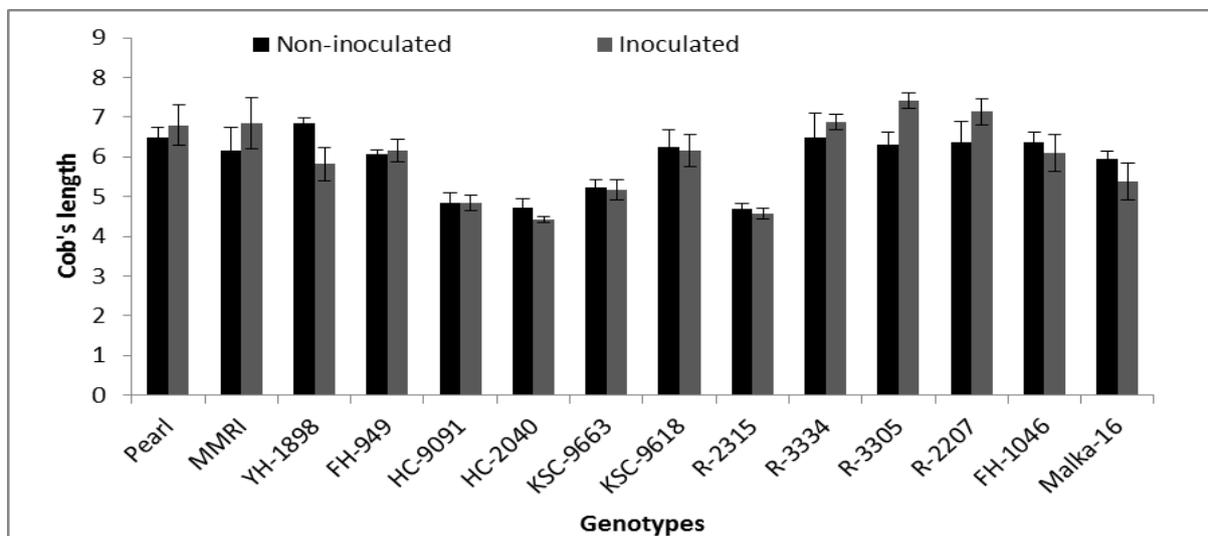


Fig. 1. Cob length (mean ± standard error SE) of different maize genotypes under *A. flavus* treatment (Least Significant Difference at 0.05 probability level = 1.12).

Analysis of variance showed 100 grains weight had highly significant difference for genotypes, treatments and Genotypes x Treatments interactions ($P < 0.001$; Table 2).

Correlation between genotypes, treatments yield related traits

Table 3 shows that there was a strong positive correlation between genotypes and treatments. Whereas, genotypes had strong negative correlation for number of grains per cob, total grains weight per cob and 100 grains weight. Treatments had strong negative correlation for number of grains per cob, total grains weight per cob and 100 grains weight.

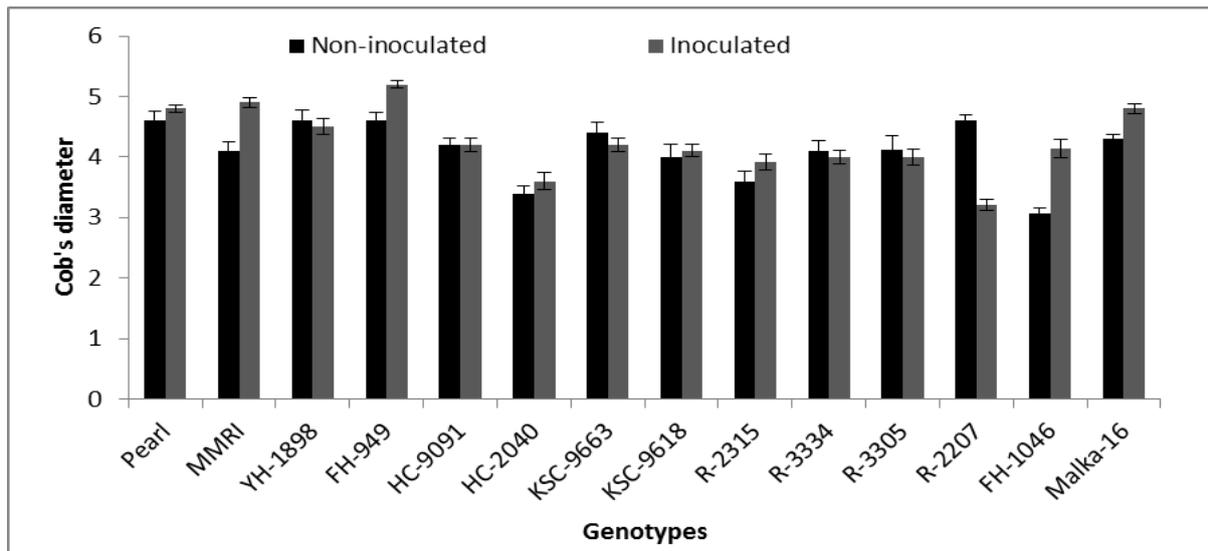


Fig. 2. Cob diameter (mean± standard error SE) of different maize genotypes under *A. flavus* treatment (Least Significant Difference at 0.05 probability level =0.42).

Cob length had strong positive correlation for cob weight, number of grains per cob and total grains weight per cob. Cob diameter had strong positive correlation for cob weight and total grains weight per cob. Cob weight had strong positive correlation for number of grains per cob, total grains weight per cob

and 100 grains weight. Number of grains per cob had positive strong correlation total grains weight per cob and weak correlation for 100 grains weight. Total grains weight per cob had strong correlation for 100 grains weight.

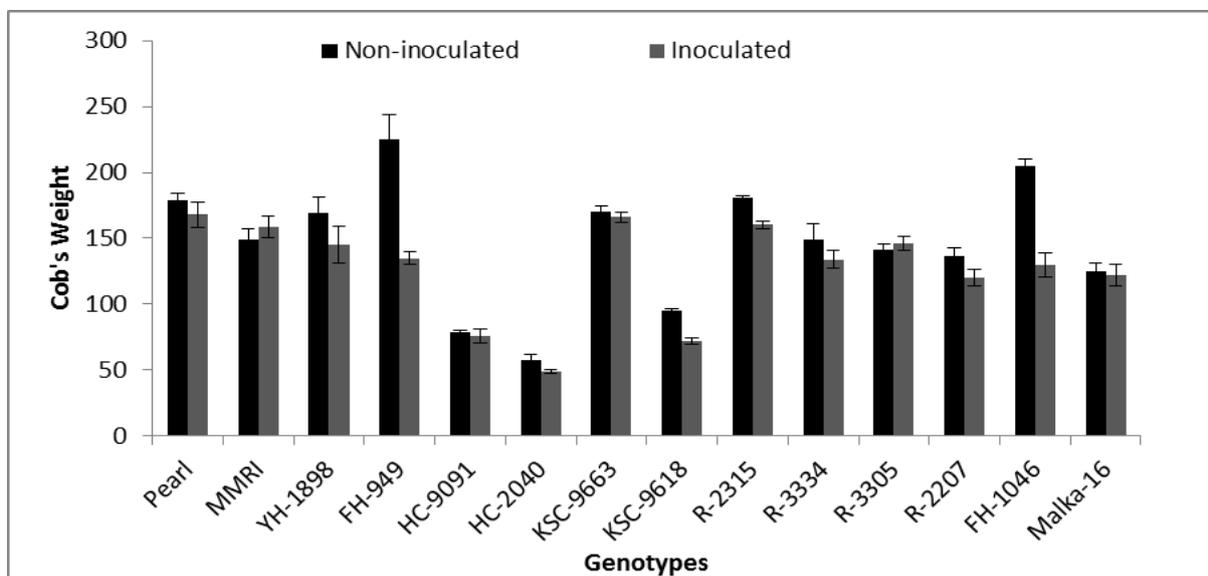


Fig. 3. Cob weight (mean± standard error SE) of different maize genotypes under *A. flavus* treatment (Least Significant Difference at 0.05 probability level =23.87).

Discussion

Meteorological parameters and A. flavus inoculation
Present study was conducted in a semi-arid conditions of Pakistan which is favorable for the growth of *A. flavus*. The present study revealed that

inoculation had a negative impact on the maize yield. This is because; the concentration of the *A. flavus* inoculation was enough for producing the *A. flavus* infection and aflatoxin in the grains as earlier described by Windham and Williams (2016). In

current study *A. flavus* treatment was applied through silk, which is very nearby to the natural infection process of *A. flavus* for production of aflatoxins. Windham and Williams, (2007) described that silk inoculation method may also be sufficient for *A. flavus* growth for production of aflatoxins, but it depend upon meteorological conditions, location and nature of maize germ plasm (Williams and Windham,

2012; Windham and Williams, 2016) and similar was observed in this study. Numerous previous studies described that *A. flavus* efficiently produced infection and ultimately aflatoxin under higher temperature (Bruns and Abbas, 2005; Campa *et al.*, 2005; Bruns and Abbas, 2006; Abbas *et al.*, 2007; Bellaloui *et al.*, 2016; Windham and Williams, 2016), similarly the current investigation had the temperature above 25°C.

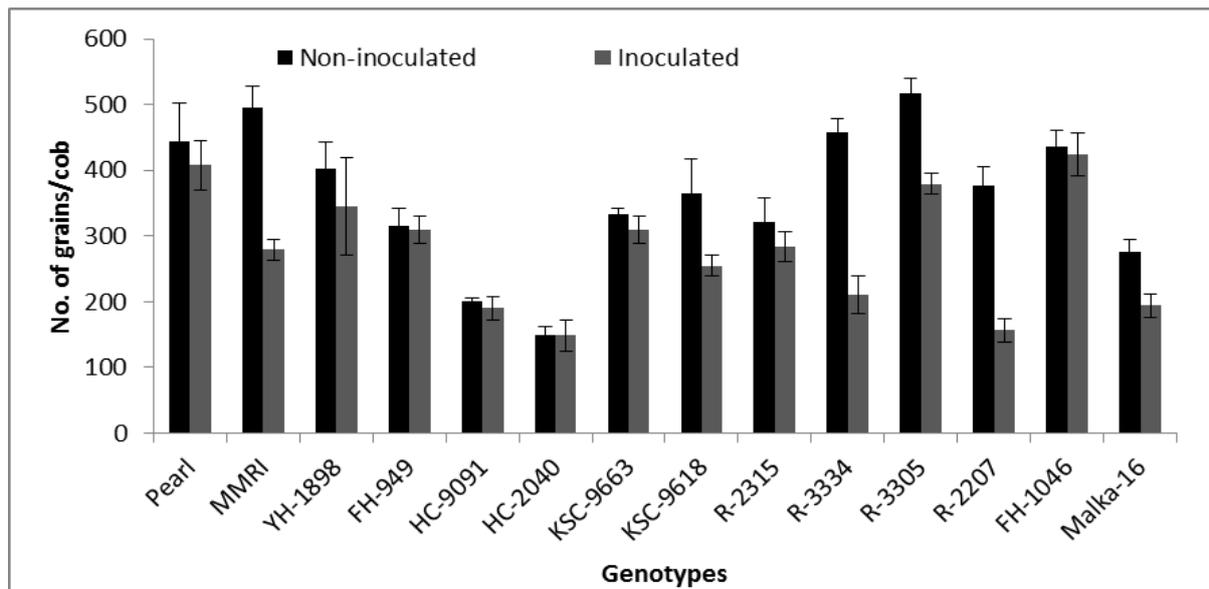


Fig. 4. No. of grains per cob (mean± standard error SE) of different maize genotypes under *A. flavus* treatment (Least Significant Difference at 0.05 probability level = 95.97).

Yield related parameters

The present study represented that *A. flavus* inoculated and non-inoculated plants showed a differential response with respect to cob length because the *A. flavus* treatment was applied during grain filling stage, when cob had achieved its length. Though, the maize genotype, HC-2040 showed relatively lower values than those of all other genotypes for almost all yield related parameters recorded during current investigation.

Maize genotype R-3305 had highest values for cob length and no. of grains per cob, whereas maize genotype FH-949 had higher values for cob diameter and cob weight as compared to all other genotypes investigated in the current study. Maize genotype FH-1046 produced higher value for total grains weight per cob and KSC-9663 produced higher values for 100 grains weight as compared to all other genotypes

investigated in the current study. This differential behavior might be due to its genetic makeup as described in earlier studies (Grzesiak *et al.*, 2007; Ali *et al.*, 2011; Ali *et al.*, 2012a; Ali *et al.*, 2012b; Ali and Ahsan, 2015).

Present investigations revealed that treatments had strong negative correlation for number of grains per cob, total grains weight per cob and 100 grains weight. On the basis of such correlation, we can investigate the relationship between inoculation and yield.

The positive correlation of cob length with cob weight, number of grains per cob and total grains weight per cob was observed during present investigation such correlation was also observed by previous studies (Afarinesh *et al.*, 2005; Wang *et al.*, 2007; Ali *et al.*, 2013). Similarly the strong positive correlation was

observed for cob diameter with cob weight and total grains weight per cob as well as cob weight had strong positive correlation for no. of grains per cob, total grains weight per cob and 100 grains weight. Such correlations were also observed by many scientists

working on *A. flavus* and plants interaction (Afarinesh *et al.*, 2005; Grzesiak *et al.*, 2007; Ali *et al.*, 2011; Ali *et al.*, 2012a; Ali *et al.*, 2012b; Ali and Ahsan, 2015).

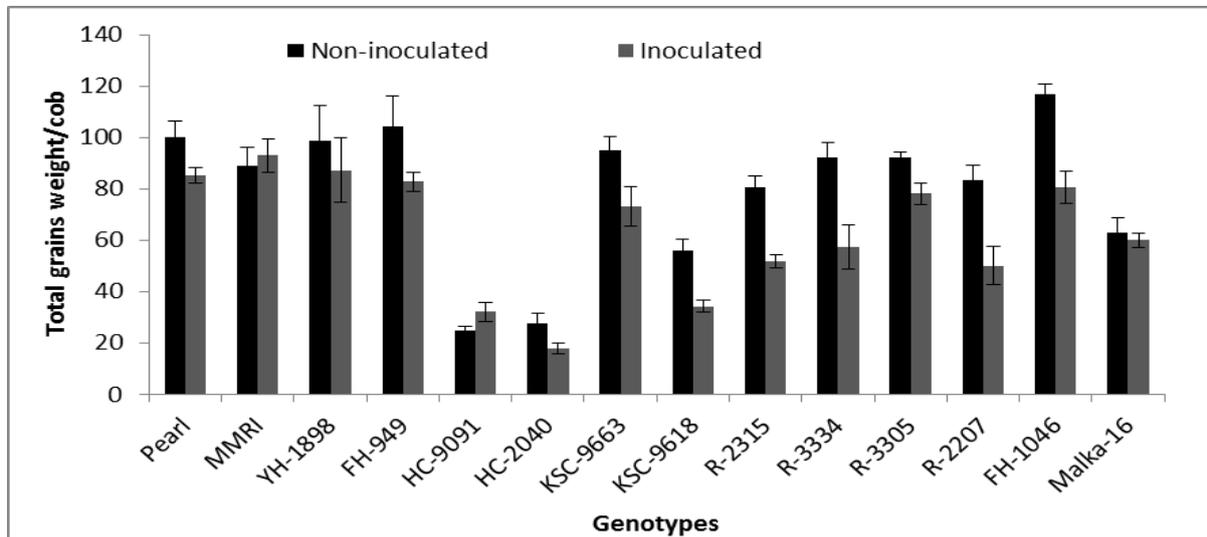


Fig. 5. Total grains weight per cob (mean± standard error SE) of different maize genotypes under *A. flavus* treatment (Least Significant Difference at 0.05 probability level = 19.74).

The findings of current investigations regarding positive correlations between yield and yield-related components of maize had also been reported by many other scientists (Ilker, 2011; Hasyan *et al.*, 2012;

Kumar *et al.*, 2014; Karasu *et al.*, 2015). However, this could be considered as selection criteria for higher grain yield potential in maize genotypes (Rani *et al.*, 2017).

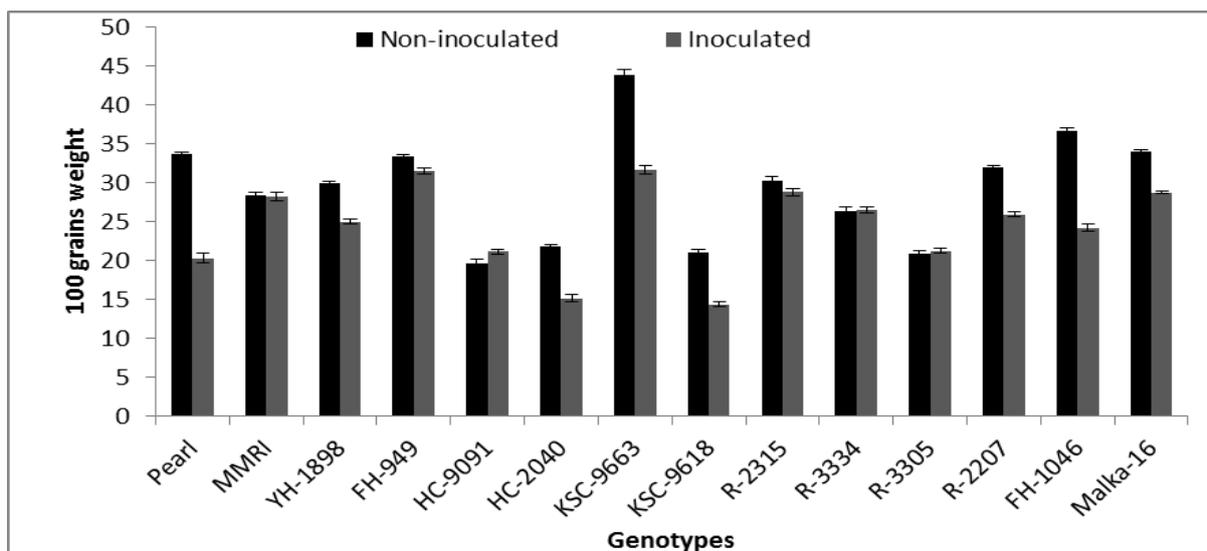


Fig. 6. 100 grains weight (mean± standard error SE) of different maize genotypes under *A. flavus* treatment (Least Significant Difference at 0.05 probability level =1.22).

Conclusion

It is concluded that *A. flavus* treatment through silk could significantly affect the yield of maize genotypes under favorable weather conditions for *A. flavus*.

References

- Abbas H, Shier W, Cartwright R.** 2007. Effect of temperature, rainfall and planting date on aflatoxin and fumonisin contamination in commercial Bt and non-Bt corn hybrids in Arkansas. *Phytoprotection* **88**, 41-50.
<http://dx.doi.org/10.7202/018054ar>
- Afarinesh A, Farshadfar E, Choukan R.** 2005. Genetic analysis of drought tolerance in maize (*Zea mays* L.) using diallel method. *Seed Plant* **20**, 457-473.
- Ali Q, Ahsan M, Ali F, Aslam M, Khan NH, Munzoor M, Mustafa HSB, Muhammad S,** 2013. Heritability, heterosis and heterobeltiosis studies for morphological traits of maize (*Zea mays*L.) seedlings. *Advancements in Life Sciences* **1**, 52-63.
- Ali Q, Ahsan M, Khan NH, Ali F, Elahi M, Elahi F.** 2012b. Genetic analysis for various quantitative traits of chickpea (*Cicer arietinum* L.). *International Journal for Agro Veterinary and Medical Sciences* **6**, 51-57.
<http://dx.doi.org/10.5455/ijavms.12877>
- Ali Q, Ahsan M, Tahir MHN, Basra SMA.** 2012a. Genetic evaluation of maize (*Zea mays* L.) accessions for growth related seedling traits. *International Journal for Agro Veterinary and Medical Sciences* **6**, 164-172.
<http://dx.doi.org/10.5455/ijavms.133>
- Ali Q, Ahsan M.** 2015. Correlation analysis for various grain contributing traits of *Zea mays*. *African Journal of Agricultural Research* **10**, 2350-2354.
<http://dx.doi.org/10.5897/AJAR2013.7838>
- Ali Q, Elahi M, Ahsan M, Tahir MHN, Basra SMA.** 2011. Genetic evaluation of maize (*Zea mays* L.) genotypes at seedling stage under moisture stress. *International Journal for Agro Veterinary and Medical Sciences* **5**, 184-193.
- Amaike S, Keller NP.** 2011. *Aspergillus flavus*. *Annual Review of Phytopathology* **49**, 107-133.
<https://doi.org/10.1146/annurev-phyto-072910-095221>
- Ashraf U, Salim MN, Sher A, Sabir SUR, Khan A, Pan SG, Tang X.** 2016. Maize growth, yield formation and water-nitrogen usage in response to varied irrigation and nitrogen supply under semi-arid climate. *Turkish Journal of Field Crops* **21(1)**, 88-96.
<http://dx.doi.org/10.17557/tjfc.93898>
- Atanda O, Ogunrinu M, Olorunfemi F.** 2011. A neutral red desiccated coconut agar for rapid detection of aflatoxigenic fungi and visual determination of aflatoxins. *World Mycotoxin Journal* **4(2)**, 147-155.
<https://doi.org/10.3920/WMJ2010.1241>
- Bellaloui N, Abbas HK, Bruns HA, Mengistu A.** 2016. Grain Chemical Composition as Affected by Genetic Backgrounds and Toxigenic *Aspergillus flavus* Inoculation in Corn Hybrids. *Atlas Journal of Plant Biology* 66-76.
<https://doi.org/10.5147/ajpb.voio.115>
- Bruns HA, Abbas HK.** 2005. Responses of short-season corn hybrids to a humid subtropical environment. *Agronomy Journal* **97**, 446-451.
<http://dx.doi.org/10.2134/agronj2005.0446>
- Bruns HA, Abbas HK.** 2006. Planting date effects on Bt and non-Bt corn in the mid-south USA. *Agronomy Journal* **98**, 100-106.
<http://dx.doi.org/10.2134/agronj2005.0143>
- Campa De La R, Hooker DC, Miller JD, Schaafsma AW, Hammond BG.** 2005. Modeling effects of environment, insect damage, and Bt genotypes on fumonisin accumulation in maize in

Argentina and the Philippines. *Mycopathologia* **159**, 539-552.

Christensen SA, Kolomiets MV. 2011. The lipid language of plant–fungal interactions. *Fungal Genetics and Biology* **48(1)**, 4-14.

<https://doi.org/10.1016/j.fgb.2010.05.005>

Farfan IDB, Gerald N, Murray SC, Isakeit T, Huang PC, Warburton, Williams P, Windham GL, Kolomiets M. 2015. Genome wide association study for drought, aflatoxin resistance, and important agronomic traits of maize hybrids in the sub-tropics. *PLoS One* **10(2)**, e0117737.

<https://doi.org/10.1371/journal.pone.0117737>

Fountain J, Scully B, Ni X, Kemerait R, Lee D, Chen ZY, Guo B. 2014. Environmental influences on maize-*Aspergillus flavus* interactions and aflatoxin production. *Frontiers in Microbiology* **5**, 40.

<https://doi.org/10.3389/fmicb.2014.00040>

Foyer CH, Noctor G. 2003. Redox sensing and signalling associated with reactive oxygen in chloroplasts, peroxisomes and mitochondria. *Physiologia Plantarum* **119(3)**, 355-364.

<https://doi.org/10.1034/j.1399-3054.2003.00223.x>

Gong M, Chen BO, Li ZG, Guo L H. 2001. Heat-shock-induced cross adaptation to heat, chilling, drought and salt stress in maize seedlings and involvement of H₂O₂. *Journal of Plant Physiology* **158(9)**, 1125-1130.

<https://doi.org/10.1078/0176-1617-00327>

Grzesiak MT, Rzepka A, Hura T, Hura K, Skoczowski A. 2007. Changes in response to drought stress of triticale and maize genotypes differing in drought tolerance. *Photosynthesis* **45**, 280-287.

Hasyan RM, Moualla YM, Ahmad AAS. 2012. Potence ratio and path coefficient analysis for some quantitative traits of maize (*Zea mays*L.) hybrids

developed in Syria. *Jordan Journal of Agricultural Sciences* **8**, 557-565.

Hiran P, Kerdchoechuen O, Laohakunjit N. 2016. Combined effects of fermentation and germination on nutritional compositions, functional properties and volatiles of maize seeds. *Journal of Cereal Science* **71**, 207-216.

<https://doi.org/10.1016/j.jcs.2016.09.001>

Ilker E. 2011. Correlation and path coefficient analyses in sweet corn. *Turkish Journal of Field Crops* **16**, 105-107.

Karasu A, Kusu H, Mehmet OZ, Bayram G. 2015. The Effect of Different Irrigation Water Levels on Grain Yield, Yield Components and Some Quality Parameters of Silage Maize (*Zea mays* in dentate Sturt.). *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* **43**, 138-145.

<http://dx.doi.org/10.15835/nbha4319602>

Kelley RY, Williams WP, Mylroie JE, Boykin DL, Harper JW, Windham GL, Ankala A, Shan X. 2012. Identification of maize genes associated with host plant resistance or susceptibility to *Aspergillus flavus* infection and aflatoxin accumulation. *PLoS One* **7(5)**, e36892.

<https://doi.org/10.1371/journal.pone.0036892>

Khan NA, Yu P, Ali M, Cone JW, Hendriks WH. 2015. Nutritive value of maize silage in relation to dairy cow performance and milk quality. *Journal of the Science of Food and Agriculture* **95(2)**, 238-252.

<https://doi.org/10.1002/jsfa.6703>

Klich MA. 2002. Identification of common *Aspergillus* species. *Centraalbureau Voor Schimmel cultures*.

Kuhnen S, Ogliari JB, Dias PF, Boffo EF, Correia I, Ferreira AG, Delgadillo I, Maraschin M. 2010. ATR-FTIR spectroscopy and chemo-metric analysis applied to discrimination of landrace maize flours produced in southern

Brazil. *International Journal of Food Science & Technology* **45(8)**, 1673-1681.

<https://doi.org/10.1111/j.1365-2621.2010.02313.x>

Kumar GP, Prashanth Y, Reddy VN, Sudheer S, Rao VP. 2014. Character association and path coefficient analysis in maize (*Zea mays* L.). *International Journal of Applied & Biology and Pharmaceutical Technology* **5**, 257-260.

Liu Y, Wu F. 2010. Global burden of aflatoxin-induced hepatocellular carcinoma: a risk assessment. *Environmental Health Perspectives* **118 (6)**, 818.

<http://dx.doi.org/10.1289/ehp.0901388>

Maina AW, Wagacha JM, Mwaura FB, Muthomi JW, Woloshuk CP. 2016. Postharvest practices of maize farmers in Kaiti District, Kenya and the impact of hermetic storage on populations of *Aspergillus* spp. and aflatoxin contamination. *Journal of Food Research* **5(6)**, 53.

Mayfield K, Betrán FJ, Isakeit T, Odvody G, Murray SC, Rooney WL, Landivar JC. 2012. Registration of maize germplasm lines Tx736, Tx739, and Tx740 for reducing pre-harvest aflatoxin accumulation. *Journal of Plant Registrations* **6(1)**, 88-94.

<http://dx.doi.org/10.3198/jpr2010.12.0675crg>

Ogbaga CC, Miller MA, Johnson GN. 2017. Fourier transform infrared spectroscopic analysis of maize (*Zea mays*) subjected to progressive drought reveals involvement of lipids, amides and carbohydrates. *African Journal of Biotechnology* **16 (18)**, 1061-1066.

<https://doi.org/10.5897/AJB2017.15918>

Pastori GM, Kiddle G, Antoniw J, Bernard S, Veljovic-Jovanovic S, Verrier PJ, Noctor G, Foyer CH. 2003. Leaf vitamin C contents modulate plant defense transcripts and regulate genes that control development through hormone signaling. *The Plant Cell* **15(4)**, 939-951.

<https://doi.org/10.1105/tpc.010538>

Patt AG, Schröter D. 2008. Perceptions of climate risk in Mozambique: implications for the success of adaptation strategies. *Global Environmental Change* **18(3)**, 458-467.

<https://doi.org/10.1016/j.gloenvcha.2008.04.002>

Probst C, Schulthess F, Cotty PJ. 2010. Impact of *Aspergillus* section *Flavi* community structure on the development of lethal levels of aflatoxins in Kenyan maize (*Zea mays*). *Journal of Applied Microbiology* **108(2)**, 600-610.

<https://doi.org/10.1111/j.1365-2672.2009.04458.x>

Rani GU, Rao VS, Ahmad ML, Rao KN. 2017. Character association and path coefficient analysis of grain yield and yield components in maize (*Zea mays* L.). *International Journal of Current Microbiology and Applied Sciences* **6**, 4044-4050.

<https://doi.org/10.20546/ijcmas.2017.612.465>

Schilling J, Vivekananda J, Khan MA, Pandey N. 2013. Vulnerability to environmental risks and effects on community resilience in mid-west Nepal and south-east Pakistan. *Environment and Natural Resources Research* **3(4)**, 27.

Shakoor MB, Nawaz R, Hussain F, Raza M, Ali S, Rizwan M, Oh SE, Ahmad S. 2017. Human health implications, risk assessment and remediation of As-contaminated water: A critical review. *Science of the Total Environment* **601**, 756-769.

<https://doi.org/10.1016/j.scitotenv.2017.05.223>

Smit B, Skinner MW. 2002. Adaptation options in agriculture to climate change: a typology. *Mitigation and Adaptation Strategies for Global Change* **7(1)**, 85-114.

Wagacha JM, Muthomi JW. 2008. Mycotoxin problem in Africa: current status, implications to food safety and health and possible management strategies. *International Journal of Food Microbiology* **124(1)**, 1-12.

<https://doi.org/10.1016/j.ijfoodmicro.2008.01.008>

Wahid A, Perveen M, Gelani S, Basra SM. 2007. Pretreatment of seed with H₂O₂ improves salt tolerance of wheat seedlings by alleviation of oxidative damage and expression of stress proteins. *Journal of Plant Physiology* **164**(3), 283-294.

<https://doi.org/10.1016/j.jplph.2006.01.005>

Wang BQ, Li ZH, Duan LS, Zhai ZX. 2007. Effect of coronatine on photosynthesis parameters and endogenous hormone contents in maize (*Zea mays* L.) seedling under drought stress. *Plant Physiology Communications* **43**, 269-272.

Williams WP, Windham GL. 2012. Registration of Mp718 and Mp719 germplasm lines of maize. *Journal of Plant Registrations* **6**, 200-202.

<http://dx.doi.org/10.3198/jpr2011.09.0489crg>

Windham GL, Williams WP. 2016. Effect of inoculum concentrations of *Aspergillus flavus* and *A. parasiticus* on aflatoxin accumulation and kernel

infection in resistant and susceptible maize hybrids. *Phytoparasitica* **44**, 333-339.

<http://dx.doi.org/10.1007/s12600-016-0523-4>

Windham GL, Williams WP. 2007. A comparison of inoculation techniques for inducing aflatoxin contamination and *Aspergillus flavus* kernel infection on corn hybrids in the field. *Phytoparasitica* **35**, 244.

Windham GL, Williams WP. 2016. Effect of inoculum concentrations of *Aspergillus flavus* and *A. parasiticus* on aflatoxin accumulation and kernel infection in resistant and susceptible maize hybrids. *Phytoparasitica* **44**, 333-339.

Wu F, Munkvold GP. 2008. Mycotoxins in ethanol co-products: modeling economic impacts on the livestock industry and management strategies. *Journal of Agricultural and Food Chemistry* **56**(11), 3900-3911.

<http://dx.doi.org/10.1021/jfo72697e>