

Journal of Biodiversity and Environmental Sciences (JBES)

ISSN: 2220-6663 (Print) 2222-3045 (Online) Vol. 5, No. 1, p. 647-654, 2014 http://www.innspub.net

RESEARCH PAPER

OPEN ACCESS

A phylogenetic analysis of Hyoscyamus L. (Solanaceae) species from Iran based on ITS and trnL-F sequence data

Shadi Hajrasouliha^{1*}, Ali Asghar Massoumi², Taher Nejad Sattari¹, Seyed Mohammad Mahdi Hamdi³, Iraj Mehregan¹

Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran

²Research Institute of Forests and Rangelands, P. O. Box; 13185-116, Tehran, Iran

³Department of Biology, Garmsar Branch, Islamic Azad University, Garmsar, Iran

Article published on July 29, 2014

Key words: Crataegus, Iran, ITS, molecular data, phylogeny.

Abstract

A phylogeny of Hyoscyamus L. (Solanaceae) that represents two different subgenera (i.e Hyoscyamus subgen. Dendrotrichon SchÖnbeck-Temesy and Hyoscyamus subgen. Hyoscyamus) based on ITS and trnL-F sequence data were studied. In this study five species of Hyoscyamus were Iran endemic. ITS and trnL-F sequence data for 18 species of this genus from Iran were studied. In addition we used 8 ITS sequence data and 13 trnL-F sequence data from genbank for another taxa in Solanaceae family. For phylogenetic analysis were used Bayesian and Maximum parsimony methods. Parsimony analyses of the ITS and trnL-F regions formed a dendrogram with strong bootstrap (ca.≥70%) that showed *Hyoscyamus* is a monophyletic group. In this study four synonymous species (H.insanus Stocks, H.rosularis SchÖnbeck-Temesy, H.orthocarpus SchÖnbeck-Temesy, H.nutans SchÖnbeck-Temesy) were stand in one clade and two similisar species H.bornmulleri Khatamsaz and H.tenuicaulis SchÖnbeck-Temesy constituted one clade. Also two species H.senecionis Willd and H.pusillus L. were formed one clade in trnL-F and combined analyses

*Corresponding Author: Shadi Hajrasouliha 🖾 shadi hajrasouliha@srbiau.ac

Introduction

The Solanacae are one of the most important family of flowering plants economically, floristically, enthobotanically and scientifically. Hyoscyamus belongs to Solanaceae that has 94 genera and 2950 species, which are mostly distributed in tropical lands of America (Mabberley, 1987; Willis, 1973). This genus belongs to subfamily solanoideae, tribe Hyoscyameae and order Solanales (Olmstead et al., 2008; TU et al., 2010; APGII, 2003). In traditional classification Solanaceae icluded two subfamily such as Solanoideae and Cestroideae (D'Arcy, 1979, 1991; Hunziker, 1979, 2001; Olmstead and Palmer, 1992). Some species of this genus such as H.niger L. is cosmopolitan (Kouhsari et al., 2006). Hyoscyamus is distributed in Turkey with six species, the former USSR with eight and Pakistan with four species (Baytop, 1978; Pojarkovae, 1955; Nasir, 1985) and has 18 species and two subgenus in Flora Iranica (SchÖnbeck-Temesy, 1972). H.bornmulleri was introduced as a new species from Shiraz (Khatamsaz, 1377). Most of the species of this genus belong to Iranian-Turanian area and some belong to European-Siberian and Sahara-Sandy. Distribution of various species of this genus in most provinces of Iran indicated that this genus is adaptable to different climates. Six species of this genus are Iran endemic and two species are plateau Iran endemic. (SchÖnbeck-Temesy, 1972). In Flora of Iran H.insanus from south, south east and center of Iran is synonymous with H.orthocarpus, H.rosularis and H.nutans but in Flora Iranica H.orthocarpus from west and south, H.nutans from south, H.rosularis from center and H.insanus from south of Iran are reported as seprate species and three species H.orthocarpus, H.rosularis and H.nutans are Iran endemic but H.insanus is plateau Iran endemic (Khatamsaz, 1377; SchÖnbeck-Temesy, 1972). We use phylogenetic analysis for survey relationship of Iranian species of *Hyoscyamus* in this study.

Material and methods

Taxon sampling

Samples were collected in the field and dried in silica

gel or taken from the herbarium material deposited at TARI (Table1). Also the list of some taxa of Solanaceae for ITS and trnL-F sequence data with genbank accession number are given in table 2.

DNA extraction, amplification and sequencing Total DNA was extracted using the DNeasy plant Mini kit (zoofa gene). Amplification of the ITS region used forward primer ITSleu1(5'-GTCCACTGAACCTTATCATTTAG-3') and reverse primer ITS4(5'-TCCTCCGCTTATTGATATGC-3') in 25 μL reactions containing the following: 12.5 μL Master mix contain of (Taq DNA polymerase, dNTP, Mgcl₂) 10.5 µL water, 1 µL DMSO, 0.5 µL forward primer, 0.5 μL reverse primer plus 1 μL DNA for each reaction. The PCR program used for ITS amplification was 97 C for 2 min followed by 30 cycles of 97 C for 1min, 50 C for 1min, 72 C for 45 sec, with a 3 sec extension per cycle and a single cycle of 72 C for 7 min (Bohs and Olmstead, 2001). Amplification of the trnL-F region used universal primers c (5'-CGAAATCGGTAGACGCTACG-3') and f (5'-ATTTGAACTGGTGACACGAG-3').

0

o

The PCR program used for trnL-F amplification was 94 C for 1min followed by35 cycles for 1min, 72 C for 2 min and a single cycle of 72 C for 5 min (Taberlet et al., 1991). Purification and sequencing of PCR products were performed in South Korea.

Sequence editing, alignment and data analysis

We edited the PCR products after sequencing with Sequencher and Bioedit and MEGA 5.1(Tamura et al., 2011). Then for aligning the sequence date we used the Mesquite (version2:75) and Mac clade (Maddison, DR and Maddison, 2000; Maddison, WP and Maddison, 2010). The matrice data were analyzed with two methods (Maximum parsimony and Bayesin) with PAUP*4.ob10 (Swofford, 2002) and MrBayes 3 (Ronquist and Huelsenbeck, 2003). In parsimony analysis all characters had equal weight that implement tree- bisection - reconnection (TBR) branch swapping with 10 heuristic random addition

648 | Hajrasouliha et al

sequence replicates. Gaps were treated as missing data. Phylogenetic analyses of 18 species of *Hyoscyamus* in two subgenus from Iran were studied. In this study we used the ITS sequence data of two species of *Anisodus*, one species of *Atropa*, one species of *Atropanthe*, three species of *Hyoscyamus*, one species of *Atropa*, two species of *Physochlaina*, one species of *Przewalskia*

and two species of Scopolia from genbank.

Results and discussion

In ITS and *trn*L-F the best modeltest of sequence evolution was modeltest 3.4 (GTR+I+G) with the following setting: nst=6, rates=gamma, base=est, rmat=est, pinv=est (Huelsenbeck and Crandall, 1997).

Table 1. List of Iranian *Hyoscyamus* species with collector name and herbarium code All collected species are saving in herbarium of research institute of forests and rangeland (TARI).

Taxon	Collector name and herbarium code	
H.nutans SchÖnbeck–Temesy	Poormirzayi & Shakeri 4968(TARI)	
H.senecionis Willd	Shams & Feizy 12828(TARI)	
H.tenuicaulis SchÖnbeck–Temesy	Assadi & Aboohamzeh 38538(TARI)	
H.orthocarpus SchÖnbeck–Temesy	Khatamsaz & Ghafari 72954(TARI)	
Hyoscyamus kurdicus Bornm	Mozaffarian 45931(TARI)	
Hyoscyamus pojarkovae SchÖnbeck–Temesy	Bazargan & Arazm 21735(TARI)	
Hyoscyamus squarrosus Griff	Assadi & Mozaffarian 35824(TARI)	
Hyoscyamus reticulatus L.	Khatamsaz & Farzaneh 73146(TARI)	
Hyoscyamus bornmulleri Khatamsaz	Mozaffarin 71291(TARI)	
Hyoscyamus rosularis SchÖnbeck–Temesy	Wendelbow & Foroughi 15292(TARI)	
Hyoscyamus malekianus Parsa	Mozaffarian 53092(TARI)	
Hyoscyamus turcomanicus Pojark	Mozaffarian 5758(TARI)	
Hyoscyamus leptocalyx Stapf	Wendelbow & Assadi 16736(TARI)	
Hyoscyamus niger L.	Massoumi & Hajrasouliha 97869(TARI)	
Hyoscyamus leucanthera Bornm & Gauba	Reshinger 50924(TARI)	
Hyoscyamus pusillus L.	Foroughi 8546(TARI)	
Hyoscyamus arachnoideus Pojark	Khatamsaz & Farzaneh 73089(TARI)	
Hyoscyamus insanus Stocks	Assadi 22750(TARI)	

ITS sequence data

The ITS data included 503 characters that 247 characters constant and 256 characters were variable. In variable characters 101 characters parsimony uninformative and 155 characters were parsimony informative.

Strict consensus tree in ITS with 520 steps was included consistency index(CI)= 0.679 and the retention index (RI) = 0.766.

All the species that we studied belong to tribe

Hyoscyameae. Within the Hyoscyameae five major clades were identified (*Atropa*, *Atropanthe*, *Anisodus*, *Scopolia* and *Hyoscyamus*). Clade *Atropa* included two samples of *Atropa belladonna* with two different genbank accession numbers (PP=1, BS=100%). *Atropanthe sinensis* constituted clade *Atropanthe* (PP=0.92, BS=100%). *Anisodus loridus* constituted *Anisodus* clade (PP=0.96, BS=100%) and *Anisodus tanguticus* was outgroup. Three species of *Scopolia* such as *Scopolia lutescens*, *Scopolia parviflora* and *Scopolia carniolica* were formed a monophyletic group (PP=1, BS=100%). And all

species of Hyoscyamus were formed a monophyletic group (18 species; PP=1, BS=100%). Hyoscyamus clade included six major clades (A-F): Clade A included four species H.insanus, H.rosularis, H.orthocarpus and *H.nutans* from subgenus Dendrotichon (PP=1, BS=100%). This clade has two subclade, one subclade included three species H.insanus, H.rosularis, H.orthocarpus (PP=0.98, BS=80%). H.untans consitituted another subclade. Clade C included two species H.bornmulleri and H.tenuicaulis from subgenus Dendrotrichon (PP=1, BS=100%). Clade F included nine species H.senecionis, H.kurdicus Bornm, H.leucanthera Bornm & Gauba, *H.pojarkovae* Schönbeck-Temesy, *H.reticulatus* L., *H.squarrosus* Griff, *H.arachnoideus* Pojark, *H.pusillus*, *H.turcomanicus* Pojark from subgenus *Hyoscyamus* (PP=0.71). This clade has two subclade (FI, FII). In subclade FII six species *H.kurdicus*, *H.leucanthera*, *H.pojarkovae*, *H.reticulatus*, *H.squarrosus* and *H.arachnoideus* were formed a polytomy group (PP=0.9). Also two species *H.pusillus* and *H.turcomanicus* were formed another subclade (PP=0.9). Finally another species of *Hyoscyamus* clade such as *H.malekianus* Parsa were stand in clade B, *H.leptocalyx* Stapf in clade D, *H.niger* in clade E and *H.senecionis* in subclade FI.

Table 2. List of some taxa in Solanaceae with genbank accession number for ITS and trnL-F sequence data.

Taxon	GenBank accession number	
_	ITS	
		trnLF
Anisodus luridus	AY478400.1_AY478414.1	
Anisodus tangiticus	AY478402.1_AY478416.1	
Atropa belladonna	AB019288_AB019948.1	
Atropa belladonna	AY028129_AY028147	
Atropanthe sinensis	AY478403.1_AY478417.1	
Scopolia carniolica	AY478397.1_AY478411.1	
Scopolia lutescens	AY478395.1_AY478409.1	
Scopolia parviflora	AY478390.1_AY478404.1	
Hyoscyamus muticus		EU58099.1
Hyoscyamus albus		EU580994.1
Hyoscyamus boveanus		EU580995.1
Hyoscyamus desertorus		EU580996.1
Hyoscyamus pusillus		EU580998.1
Anisodus tanguticus		EU580956.1
Atropa belladonna		EU580958.1
Atropa belladonna		EU580959.1
Physochlaina infundibularis		EU581046.1
Physochlaina orientalis		EU581047.1
Przewalskia tangutica		EU581050.1
Scopolia carniolica		EU581060.1
Scopolia japonica		EU681061.1

TrnL-F sequence data

The *trn*L-F data included 930 characters that 854 characters constant and 76 characters were variable. In variable characters 39 characters parsimony uninformative and 37 characters were parsimony informative. Strict consensus tree in *trn*L-F with 84

steps was included (CI)= 0.940 and (RI) = 0.970.

We studied 31 samples of Solanaceae from six different genus (*Atropa, Physochlaina, Przewalskia, Scopolia, Hyoscyamus, Anisodus*) in *trn*L-F analysis. All species of *Hyoscyamus* were formed a monophyletic group (21species, PP=1, BS=100%).

Whithin this group two major clade were identified (A - B). H.muticus constituted clade A. Clade B divided to five subclade (B1-B2-B3-B4 and B5). H.desertorus constituted clade B1. Clade B2 included eight species H.boveanus. H.malekianus, H.bornmulleri, H.tenuicaulis, H.insanus, H.orthocarpus, H.rosularis and H.nutans (PP=0.97). In this clade two species H.bornmulleri and H.tenuicaulis were formed one subclade (PP=0.99, BS=70%) and four species H.insanus, H.orthocarpus, H.rosularis and H.nutans were formed another subclade (PP=0.99, BS=100%). H.leptocalyx constituted clade B3. H.turcomanicus constituted clade B4. Clade B5 included ten species (PP=0.79, BS=73%) from subgenus Hyoscyamus and divided to two subclade (B5a - B5b). Eight species H.albus, H.leucanthera, H.niger, H.pojarkovae, H.reticulatus, H.arachnoideus, H.squarrosus and H.kurdicus were formed polytomy group (subclade B5a, PP=1, BS=90%). And two species H.senecionis and H.pusillus were formed subclade B5b (PP=1, BS=100%). Among another taxa from Solanaceae we studied two samples of Atropa belladonna with two different genbank accession numbers that constituted one clade (PP=1, BS=100%). Five species such as Physochlaina Physochlaina orientalis, infundibularis, Przewalskia tangutica, Scopolia japonica and Scopolia carniolica were formed one clade (PP=0.95). This clade divided to three subclade. Two species Physochlaina orientalis and Physochlaina infundibularis were formed monophyletic group(PP=0.85, BS=75%). Two species Przewalskia tangutica and Scopolia japonica were formed one subclade (PP=0.83, BS=70%) and Scopolia carniolica constituted another subclade and Anisodus tanguticus was outgroup.

Combined sequence data

In combined analysis the best fit model was (HKY+I+G) that selected by hLRT in model test 3.7 (Posada and Crandall, 1998). The base frequencies were freq A = 0.2867, freq C= 0.2328, freq G= 0.2452 and freq T= 0.2353.

The combined data included 1391 characters that 1117 characters constant and 274 characters were variable. In variable characters 139 characters parsimony uninformative and 135 characters were parsimony informative.

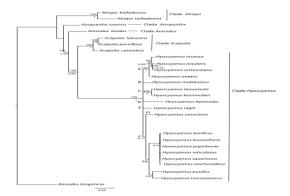


Fig. 1. The 50% majority rule consensus tree from Bayesian analysis of NrDNA ITS sequence data. Numbers above branches represent Boot strap support (BS) and numbers below branches represent posterior probabilities(PP).

Strict consensus tree in combined with 456 steps was included CI= 0.764 and RI= 0.724. homogeneity test (HI) = 0.1100.

In combined analysis all species of *Hyoscyamus* were formed a monophyletic group (18 species, PP=1, BS=100%). Within this group two major clade were indentified (A-B): H.leptocalyx constituted clade B. Clade A divided to four subclade (PP=0.91, BS=71%) (A1-A2-A3-A4): Clade A1 included two subclade A1a, A1b (BS=84%). H.turcomanicus constituted A1b. In clade A1a two species H.pusillus and H.senecionis were formed one clade (PP=1, BS=84%). Seven species H.arachnoideus, H.kurdicus, H.leucanthera, H.pojarkovae, H.squarrosus, H.reticulatus and H.niger were formed another clade (PP=1, BS=93%) In this clade H.niger constituted one subclade and another six species were formed one subclade (PP=1, BS=74%) that in this subclade four species H.kurdicus, H.leucanthera, H.pojarkovae, H.squarrosus were formed a polytomy group (PP=0.99, BS=80%). Clade A2 included H.tenuicaulis and H.bornmulleri (PP=1, BS=100). Clade A3 included H.insanus, H.orthocarpus, H.rosularis and H.nutans (PP=1, BS=100%). In this clade three species H.insanus, H.orthocarpus and H.rosularis were formed one subclade (PP=0.99, BS=73%) and H.malekianus constituted clade A4. H.insanus, H.orthocarpus, H.rosularis and H.nutans reported as a synonymous in Flora of Iran and were located in subgenus Dendrotrichon. All phylogenetic trees resulted from MP and Bayesian analyses based on ITS, trnL-F and combined sequences data showed that these four species were formed one clade and MP supported this result with strong bootstrap 100%. Then the result of present study were confirmed by Flora of Iran (Khatamsaz, 1377). Also two species H.bornmulleri and H.tenuicaulis that morphological similarity were located in one clade and belong to subgenus Dendrotrichon that placed at the closer distance from another taxa of this subgenus in trnL-F and combined analysis. H.pusillus and H.senecionis in two analyses trnL-F (BS=100%) and combined (BS=84%) were formed one clade.

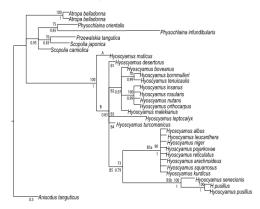


Fig. 2. The 50% majority rule consensus tree from Bayesian analysis of trnL-F sequence data. Numbers above branches represent Boot strap support (BS) and numbers below branches represent posterior probabilities(PP).

Olmstead et al (2008) studied on 89 genera and 190 species from Solanaceae by using ndhF and trnL-F markers and reported that Hyoscyamus was a monophyletic group. Also two species Physochlaina orientalis and Physochlaina infundibularis were formed a monophyletic group and two species Scopolia japonica and Przewalskia tangutica were formed a one clade that agreement with our resulst.

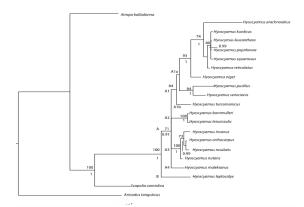


Fig. 3. The 50% majority rule consensus tree from Bayesian analysis of combined sequence data. Numbers above branches represent Boot strap support (BS) and numbers below branches represent posterior probabilities(PP).

Acknowledgements

The authors wish to thank the Islamic Azad University, Science and Research Branch Tehran for providing the facilities necessary to carry out the work and the colleagues in the herbaria TARI for providing herbarium materials.

References

APGII(Angiosperm phylogeny group) 2003. An update for the angiosperm phylogeny group classification for the orders and families of flowering plants: APGII Botanical Journal of the Linnean Society 141(4), 399-436.

http://dx.doi.org/10.1046/j.1095-8339.2003.t01-1-00158.x

Baytop A. 1978. Flora of Turkey and the East Aegean Island 6, Edinburgh at the university press. http://dx.doi.org/10.3417/2008110

Bohs L, Olmstead RG. 2001. A reassessment of Normania and Triguera (Solanaceae) 228, 33-48.

http://dx.doi.org/10.1007/s006060170035

D'Arcy WG. 1979. The classification of the Solanaceae P.p. 3-48 in: Hawkes JG, Lester RN, Skelding AD, (eds.), The Biology and Taxonomy of the Solanaceae. Academic press, London.

D'Arcy WG. 1991. The Solanaceae since 1976, with review of its biogeography. 75-139 in: Hawkes JG, Lester RN, Nee M, Strada RN, (eds.), Solanaceae III: Taxonomy, Chemistry, Evolution. Royal Botanical Garrdens, Kew.

Huelsenbeck John P, Crandall Keith A. 1997. Phylogeny estimation and hypothesis testing using maximum likelihood, Annual Review of Ecology and Systematics 28, 437-466.

http://dx.doi.org/10.1146/annurev.ecolsys.28.1.437

Hunziker AT. 1979. South American Solanaceae: a synoptic survey. P.p. 49-86 in Hawkes JG, Lester RN, Skelding AD, (eds.), The Biology and Taxonomy of the Solanaceae. Academic press London.

Hunziker AT. 2001.Genera Solanacearum.The genera of Solanaceae illustrated, arranged according to a New system Gantner, Ruggell (Liechtenstein). http://dx.doi.org/103417/2009019

Khatamsaz M. 1377. Flora of Iran, Solanaceae family research institute of forests and rangelands Vol

http://dx.doi.org/10.12692/ijb/4.6.6-18

Kouhsari SHM, Sharifi G, Ebrahimzadeh H, Khatamsaz M. 2006. comprative study of six isoenzyme systems in some species of Hyoscyamus L. from Iran, Pakistan journal of Botany 38, 107-119.

Mabberley DI. 1987. The plant book Cambridge University press, cambridge, New York. http://dx.doi.org/10.1126/science.1166605

Maddison DR, Maddison WP. 2000. Macclade4: Analysis of phylogeny and character Evolution-Sinauer Associates.

http://dx.doi.org/10.1600/036364407780360210

Maddison WP, Maddison DR. 2010. Mesquite (version2.75): A modular system for evolutionary analysis.

http://dx.doi.org/10.1098/rspb.2010.2220.6

Nasir YJ. 1985. Flora of Pakistan, Solanaseae Department of botany, university of krachi, krachi 168.

Olmstead RG, Bohs L, Abdel-Migid H, Santiago-Valentin E, Garcia VF, Collier SM. 2008. A molecular phylogeny of the Solanaceae, Taxon: 1159-1181

http://dx.doi.org/10.11646/phytotaxa.116.2.1

Olmstead RG, Palmer JD. 1992. A chloroplast DNA phylogeny of the Solanaceae: subfamilial relationships and character evolution. Ann Missouri Botanical Garden 79, 346-360.

http://dx.doi.org/10.2307/2666726

Pojarkovae AI. 1955. in schischkin BK, Bobrov EG (eds), Flora SSSR, Tom XXII Akademiya Nauk SSSR publishers, Moscow-Leningrad.

Posada D, Crandall KA. 1998. Modeltest: testing the model of DNA substitution, Bioinformatics, **14(9)**, 817-818.

http://dx.doi.org/10.1093/bioinformatics/14.9.817

Ronquist F, Huelsenbeck JP. 2003. MrBayes 3. Bayesian phylogenetic inference under mixed models, Bioinformatics 19(12), 1572-1574.

http://dx.doi.org/10.1093/bioinformatics/btg180

SchÖnbeck-Temesy E. 1972. Solanaceae In Rechinger KH (ed), Flora Iranica, Vol100, akademische druck and verlagsanstalt Granz 1, 12-72.

Swofford DL. 2002. PAUP*: Phylogenetic analysis using parsimony (*and other methods), version 40b10. Sunderland, Sinauer Associates.

http://dx.doi.org/10.1115/2009/897364

Taberlet Pierre, Gielly Ludovic, Pautou Guy, Bouvet Jean. 1991. Universal primers amplification of three non-coding regions of chloroplast DNA. Plant Molecular Biology 17, 1105-1109.

http://dx.doi.org/10.1007/BF00037152

Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. 2011. MEGA 5, Molecular genetics analysis using Maximum likelihood, Evolutionary distance.

http://dx.doi.org/10.1093/molbev/msr121

Tu T, Volis S, Dillon MO, Sun H, Wen J. 2010.

Dispersals of Hyoscyameae and Mandragoreae (solanaceae) from the New world to Eurasia in the early miocene and their biogeographic diversification within Eurasia, Molecular Phylogenetics and Evolution **57(3)**, 1225-1237.

http://dx.doi.org/10.1016/j.ympev.2010.09.007

Willis JC. 1973. A dictionary of the flowering plants and ferns VII ed, University press, Cambridge.