



Seed germination in response to osmotic stress

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Abstract

An Attempt was carried out to evaluate seed germination performances of Baraka, Adlib and Nineveh lentil cultivars besides Local Vetch and Local Mungbean cultivar under (0, -0.5, -1 and -1.5 Mpa) osmotic potentials created by dissolving pure NaCl in distilled water. Gradual reductions in osmotic solutions resulted in gradual reduction in all detected parameters. Subsequently, -1.5Mpa revealed the highest reductions in terms of final germination percentage (467.1%), germination rate (1710%), radical length (12829.4%) and Plumule length (infinity). It also aggravated the adverse effects by increasing the duration required for attaining the peak germination percentage (110.8%), as compared to that of distilled water. Treatments were categorized according to their adverse influence on performance of seed germinations as following: -1.5 Mpa> -1 Mpa> -0.5 Mpa> 0 Mpa. Mungbeans local cultivar seeds revealed the best germination performance, as compared to other pulse crops and their cultivars. Since this cultivar gave the highest germination rate (60.5 seedling.d⁻¹), plumule length (33 mm). In addition to that it significantly reduced days required for peak germination (2.6) and days to emergence commencements (1.3). Therefore, cultivars can be grouped according to their positively performance as below: Mungbean> Adlib>Nineveh> Baraka> Common Vetch. Mungbeans seeds appeared to be the most potent under all tested osmotic potentials. This cultivar manifested the highest plumule lengths under distilled water (108 mm), -0.5 Mpa (21mm) and -1.5 Mpa (3mm). Moreover this cultivar exhibited, days required for first emergence at all osmotic potentials.

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Introduction

Salt tolerance mechanism mainly preponderance by means that capable to excludes Na^+ from being in contact with functional enzymes to ensure enzyme inactivation. This task could be fulfilled by vast of gene expression, generate many enzymes responsible for transporting, sequestering and or secreting sodium ions throughout tissues. Glenn *et al.* (1999). Inge *et al.* (2009) postulated that modification of specific root Na^+ transport processes might improve Na^+ exclusion from the shoot and result, at least for some plants, in an increase in salinity tolerance. For example, initial influx of Na^+ from the soil could be decreased in the outer cell layers of the root, or efflux of Na^+ from these cells to the apoplast or soil solution could be increased. In the stele cells surrounding the vasculature, the loading of Na^+ into the xylem vessels could be decreased or retrieval of Na^+ from the transpiration stream increased. Accordingly, at the cellular level, Na^+ transport processes need to be modified in opposite directions in the inner and outer parts of the root to minimize Na^+ accumulation in the shoot. Consequently, plasma membrane Na^+ transport processes in the root need to be altered in a cell type-specific manner. Omami (2005) stated that under high salt concentration, plants sequester more NaCl in the leaf tissue than normally occurs. Increases in NaCl within the leaf tissue then result in lower osmotic potentials and more negative water potentials.

Under saline conditions, the osmotic adjustment, which occurs through the accumulation of inorganic compounds (mainly Na^+ and Cl^-) in plant, is less energy and carbon demanding than adjustment by organic solutes (Greenway and Munns, 1983). Maintenance of adequate levels of K^+ is essential for plant survival in saline habitats. Potassium is the most prominent inorganic plant solute, and as such makes a major contribution to the low osmotic potential in the stele of the roots that is a prerequisite

for turgor pressure driven solute transport in the xylem and the water balance of plants (Marschner, 1995).

Water stress is usually created from water conductance constraints namely high osmoticity at the rizosphere, root absorption barriers, vessel conduit capability and stomata behaviours. Omami (2005) reported that the reduction in root hydraulic conductance reduces the amount of water flow from the roots to the upper portion of the canopy, causing water stress in the leaf tissue. However, (Shalhevet and Hsiao, 1986) found that the growth rate under water stress was half as large as under salt stress in the leaf water potential range of interest. Nevertheless, the deleterious effects of salinity on plant growth are associated with (1) low osmotic potential of soil solution (water stress), (2) nutritional imbalance, (3) specific ion effect (salt stress), or (4) a combination of these factors (Marschner, 1995). Sohan *et al.* (1999) revealed that osmotic effects of salt on plants are as a result of a lowering of the soil water potential due to increasing solute concentration in the root zone. Therefore, at very low soil water potentials, this condition interferes with plants ability to extract water from the soil and maintain turgour. Reduction of water uptake with salinity could be related to reductions in morphological and/or physiological parameters like leaf area, stomata density, and stomata closure (stomata conductance and transpiration). Since response to saline water varies greatly with species or cultivar (Bayuelo-Jiménez *et al.*, 2003).

Above 100 mM NaCl, the delay in the onset of germination was accompanied by reductions in the final germination percentage which decreased as the NaCl concentration increased. At NaCl concentrations of 200 mM and above, no germination was observed within 72 hrs of the start of imbibitions (Scorer *et al.*, 1985). They observed

that NaCl greatly reduced the germination response of the seeds to R. The decreased R sensitivity observed in NaCl stressed seeds compares the influence response curves obtained with seeds germinated in water, 50 and 100 mM NaCl.

Germination tests were conducted under five osmotic potential levels (-0.45, -0.77, -1.03, -1.44 MPa, and Control) of PEG 6000 and NaCl. Germination percentage (%) at 4 and 8th days and also seedling growth traits such as root and shoot length (mm), dry root and shoot weight (mg), root: shoot length (R: S) ratio, and relative water content of shoot (RWC, %) were investigated in this study (Kaydan and Yagmur, 2008). Their results indicated that decreases in the osmotic potentials caused a reduction in germination percentage and seedling growth. Seed germination completed in all seed size under control solution and at -0.45 MPa of NaCl at the 8th day. Although, medium and small seeds had low germination percentage at the -0.77 MPa of NaCl, all large seeds germinated at the equivalent osmotic potential. However, subsequent low osmotic potentials of NaCl decreased germination percentage. Therefore, low germination percentage recorded at the highest NaCl concentration in all seed size. The objective of this investigation was to determine the germination performance of mungbeans, common vetch and three lentil cultivars under varying salt rates.

Materials and methods

This investigation was fulfilled at the laboratory of Field Crops Department, College of Agriculture, Salahalddin University, Erbil, Kurdistan Region, Iraq.

Factorial Randomized Complete Block Design was used in this experiment where factor (A) contained four osmotic potentials (0 Mpa (a₁), -0.5 Mpa (a₂), -1 Mpa (a₃, and -1.5 Mpa (a₄). Whereas factor (B) was represented by Nineveh lentil cv. (b₁), Adlib lentil cv. (b₂), Baraka lentil cv. (b₃), Local Common Vetch cv.

(b₄) and Local Mungbean cv. (b₅). Subsequently, 20 treatments were included in this investigation. Every treatment was replicated 4 times and one replicate contained 4 plastic disposable dishes each of 25 seeds.

NaCl solutions was detected by electrical conductivity device and osmotic potential of the prepared solutions were calculated from Ayers and Wescot (1985) equation (Osmotic potential = - 0.36 × ECe). 25 seeds were laid uniformly over salt wetted Watmann filter paper and sealed by polyethylene sheets to avoid seed desiccations. Germinated seeds were daily counted. Duration required for peak germination (days), and days for emergence commencements were counted. Final germination percentage, germination energy percentage were calculated from dividing number of germinated seeds on total seeds and from yield of number of germinated seeds after three days from starting divided on the total seeds, respectively, (Ruan *et al.*, 2002). Germination rate: germination percentage ratio was calculated from dividing the Germination rate over germination percentage. Radical and plumule lengths (mm) were measured by mini roller.

Germination rate (seedling.d⁻¹) was calculated from the following formula (Carleton, 1968): SG = No. of grains emerged at first count / Days of first count + ...+ No. of grains emerged at final count / days of final count. Mean germination time (days) was calculated from the equation below:

$$(MGR = \frac{\sum nidi}{N}); \text{ where } ni = \text{ number of}$$

germinated seeds on day I, d= rank order of day i (number of days counted from the beginning of germinated), and N=total number of germinated seeds. Finally, data were analyzed by computer statistical program, using Duncan's Multiple Range Test at $\alpha = 0.05$ probability level. Finally permanent

slides were prepared with some modification to that reported by Berlyn and Mksche (1976).

Results

Influence of NaCl concentrations

Germination of seeds under -1.5 Mpa (Table 1 and Fig. 1,a,b,c) profoundly inferior in all detected parameters, as compared to seeds germinating under distilled water (0Mpa) in terms of final germination percentage (467.1%), mean germination time (143%), germination energy (9300%), germination rate (1710%), ratio of germination rate to germination percentage (58.22%), radical length (12829.4%) and Plumule length (infinity). It also aggravated the

adverse effects by increasing the duration required for attaining the peak germination percentage (110.8%), days required for first emergence (211.1%). When this treatment was compared with that of -0.5Mpa it also revealed substantially lower values in final germination percentage (438.5%), mean germination time (74.5%), germination energy (7925%), germination rate (1359.9%), ratio of germination rate: germination percentage (32%), radical length (1870.59%) and Plumule length (infinity). Additionally, this treatment revealed profound efficacies in increasing the period required for peak germination (60.8%) and days for first emergence (211%).

Table 1. Seed germination and seedling performances of Nineveh, Adlib, and Baraka lentil cultivars, Common Vetch and Mungbean in response to four osmotic potentials using NaCl Concentrations.

Treatments		Final Germination %	Mean Germination Time (days)	Germination Energy (%)	Germination Rate (seedling/day)	Days for Peak Germination
Osmotic Potential	0 Mpa	99.25a	1.665a	94.00a	56.40a	3.700d
	-0.5 Mpa	94.25b	1.195b	80.25b	45.478b	4.850c
	-1.0 Mpa	78.25c	1.283b	27.15c	23.473c	6.750b
	-1.5 Mpa	17.5d	0.685c	1.000d	3.115d	7.800a
Legume Crops	N	72.188a	1.356a	49.5b	26.963b	6.938a
	A	74.375a	1.316a	51.313b	27.325b	6.25b
	B	72.188a	1.278a	47.5c	24.988c	6.188b
	Common Vetch	69.688b	1.078b	35.0d	20.844d	6.938a
0 Mpa	Mungbean	73.125a	1.047b	69.688a	60.463a	2.563c
	N	97.5a	1.938a	93.75bc	47.425c	4.75d
	A	100.0a	1.438b	83.75de	38.6de	4.0de
	B	98.75a	1.025def	100.0a	100.0a	2.0f
-0.5 Mpa	Common Vetch	100a	1.563b	87.5d	38.1de	7.5b
	Mungbean	100a	1.325bcd	92.5c	36.725e	4.0de
	N	97.5a	1.063cf	80.0e	31.175f	4.75d
	A	92.5b	0.987f	41.25g	27.225g	6.0c
-1.0 Mpa	B	88.75bc	1.088cf	100.0a	94.165b	2.0f
	Common Vetch	92.5b	1.35bc	16.75b	21.025h	7.5b
	Mungbean	100 a	1.363bc	15.25h	21.275h	8.0b
	N	83.75d	1.263be	15.0h	19.575h	7.75b
-1.5 Mpa	A	86.25cd	0.975ef	15.0h	13.80i	8.5ab
	B	83.75 d	1.463b	73.75f	41.688d	2.0f

	Common Vetch	62.5f	0.575g	0.0j	1.30k	8.0b
	Mungbean	75e	0.625g	0.0j	2.375jk	9.5a
-1.5 Mpa	N	10i	0.65g	0.0j	2.15k	8.0b
	A	18.75h	0.963ef	0.0j	3.75jk	9.25a
	B	17.5h	0.613g	5.0i	6.0j	4.25de
	Common Vetch	23.75g	0.963ef	0.0j	3.75jk	9.25a
	Mungbean	17.5h	0.613g	5.0i	6.0j	4.25de
	Treatments	Days for First Emergence	Germination Rate: Germination % Ratio	Radical Length (mm)	Plumule Length (mm)	
Osmotic Potential	0 Mpa	1.80c	0.568a	109.9a	62.35a	
	-0.5 Mpa	1.80c	0.474b	16.75b	10.0b	
	-1.0 Mpa	2.6b	0.31d	2.35c	1.9c	
	-1.5 Mpa	5.6a	0.359c	0.85b	0.00d	
Legume Crops	N	3.5a	0.311c	34.656b	15.875bc	
	A	3.375ab	0.315c	36.625a	17.219b	
	B	3.25b	0.313c	34.531b	14.656c	
	Common Vetch	3.313b	0.49b	26.563d	12.063d	
	Mungbean	1.313c	0.718a	30.0c	33.0a	
0 Mpa	N	2.0e	0.485e	117.5b	52.5c	
	A	2.0e	0.49e	121.25a	57.5b	
	B	2.0e	0.475e	113.75c	51.25c	
	Common Vetch	2.0e	0.388fg	88.75d	42.5d	
	Mungbean	1.0f	1.0a	108.5d	108a	
-0.5 Mpa	N	2.0e	0.378fg	18.0g	10.0f	
	A	2.0e	0.398f	21.25f	9.75f	
	B	2.0e	0.355g	20.0fg	5.5g	
	Common Vetch	2.0e	0.295h	13h	3.75gh	
	Mungbean	1.0f	0.945c	11.5h	21.0e	
-1.0 Mpa	N	3.0d	0.253i	2.625ij	1.0hi	
	A	3.0d	0.245i	3.0i	1.625hi	
	B	3.0d	0.26hi	2.875ij	1.875hi	
	Common Vetch	3.0d	0.22i	3.25i	2.0hi	
	Mungbean	1.0f	0.573d	1.0k	3.0ghi	
-1.5 Mpa	N	7.0a	0.13j	0.5k	0.0i	
	A	6.5b	0.127j	1.0ijk	0.0i	
	B	6.0c	0.123j	1.5ijk	0.0i	
	Common Vetch	6.25bc	1.085a	1.25ijk	0.0i	
	Mungbean	2.25e	0.355g	0.0k	0.0i	

Table 2. Regression analysis results for the responses of germination performance to varying osmotic potential levels.

Character	Regression equation	(R ²)
Final Germination Percentage (%)	$Y = 99.25 - 21 X + 45.5 X^2 - 45 X^3$	96.7
Mean Germination Time (days)	$Y = 1.665 - 2.326 X + 3.600 X^2 - 1.657 X^3$	58.1
Germination Energy (%)	$Y = 94 + 56.05 - 211.3 X^2 + 88.400 X^3$	84.9
Germination Rate (seedling/day)	$Y = 59.396 - 36.372 X$	57.2
Days for Peak Germination	$Y = 3.645 + 2.840 X$	39.7
Days for First Emergence	$Y = 1.120 + 2.440 X$	54
Germ. Rate:Germination Percentage Ratio	$Y = 0.546 - 0.158 X$	10.6
RadicalLength (mm)	$Y = 106.655 - 205.89 X$	95.5
Plumule Length (mm)	$Y = 62.35 - 174.317 + 164.6 X^2 - 50.733 X^3$	81.5

This treatment was followed by -1Mpa in sequence order, since the latter treatment significantly reduced the final germination percentage (26.8%), mean germination duration (29.8%), germination energy (246.2%), germination rate (140.3%), germination rate : germination percentage ratio (83.2%), radical length (4576.6%), and plumule length (3181.6%). This treatment also took similar trends in increasing the duration required for peak germination (82.4%) and days for first emergence (44.4%), as compared to treatment of distilled water media. The compression between -1Mpa to that of -0.5Mpa in term of final germination percentage (20.4%), mean germination duration (7.4%), germination energy (195.6%), germination rate (93.7%), germination rate: germination percentage ratio (52.9%), radical length (612.8%), and plumule length (426.3%). It highly increased the time required for peak germination (39.2%), days required for first emergence (44.44%).

Performance of seed germinations in -0.5Mpa manifested substantial reduction in relation to germinations performed under 0 Mpa in the final germination percentage (39.3%), germination energy (17.1%), germination rates (24%), germination rate: germination percentage ratio (19.8%), radical length (556.1%), and plumule length 523.5%). Moreover, it extended the period required for peak germination (31.1%). Subsequently, the best seed germination performance was obtained from seeds germinated in distilled water. These results suggested that germination of legume seeds under solutions higher

than -0.5Mpa are not recommended owing to the risk of poor germination and low radical growth.



Fig. 1. Nature of germination and seedling performances of Mungbean in response to four osmotic potentials using NaCl concentrations.

Very close results were found by (Abdel, 1989). He germinated onion seeds in NaCl solutions at rates of 0, -5, -10 and -15 bars. Time required to first emergence, time to peak germinations, peak germination percentage, final seed germination, percentage of survived seeds after salt washing from un-germinated seeds revealed gradual substantial reduction confined with the gradual reductions in solute osmosity. These results were attributed to Na⁺ and Cl⁻ toxic effects besides water imbibitions constraints. Fenugreek seeds germination capacity in varying NaCl solutions were highly reduced particularly under -1.5 MPa in compassion to distilled water check. Reductions were in terms of peak germination percentage (92%), and final

germination percentage (94%). Salts influence on seed germination were attributed to the toxic effects of Na⁺ and Cl⁻ preponderances in cellular membrane and cytosol by which enzymes are denaturalized. Iyengar and Reddy (1996) found that salt toxicity caused particularly by Na⁺ and Cl⁻ ions; and soil salinity represents an increasing threat to agricultural production. High sodium (Na⁺) concentrations in soils are toxic to higher plants. More than 40% of irrigated lands worldwide show increased salt levels (Horie and Schroeder, 2004).

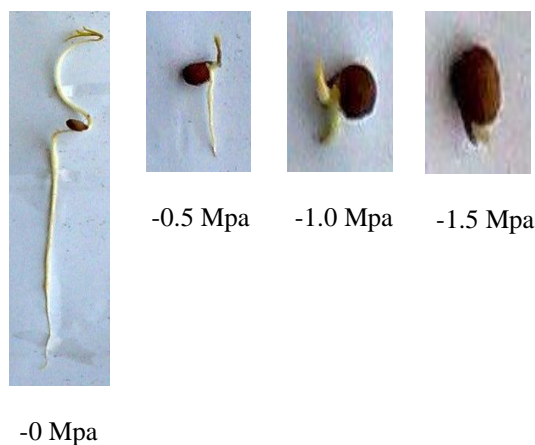


Fig. 2. Nature of germination and seedling performances of Common Vetch in response to four osmotic potentials using NaCl concentrations.

Cultivar responses

The obtained results (Table, 1 and Figure, 1, a,b,c) manifested that Mungbaen local cultivar seeds revealed the best germination performance, as compared to other pulse crops and their cultivars. Since this cultivar gave the highest germination energy (69.7%), germination rate (60.5seedling.d⁻¹), germination rate: germination percentage ratio (0.72), and plumule length (33 mm). In addition to that it significantly reduced days required for peak germination (2.6) and days to emergence commencements (1.3). Adlib lentil cultivar came next to local Mungbean in the superiority order. This cultivar was preponderated in germination energy

(51.3%), germination rate (27.3seedling.d⁻¹), and plumule length (17.2 mm). Non- significant differences were observed between Adlib and Mungbean in final germination percentage, besides its overwhelming over all detected cultivars in radical length (36.5mm). The third cultivar in the sequence order was Nineveh cultivar which substantially exceeded Braka and Common Vetch in germination energy (4.2% and 41.4%, respectively) and germination rate (7.9% and 29.4%, respectively) and it highly exceeded Common Vetch in both radical length (30.5%) and plumule length (31.6%).

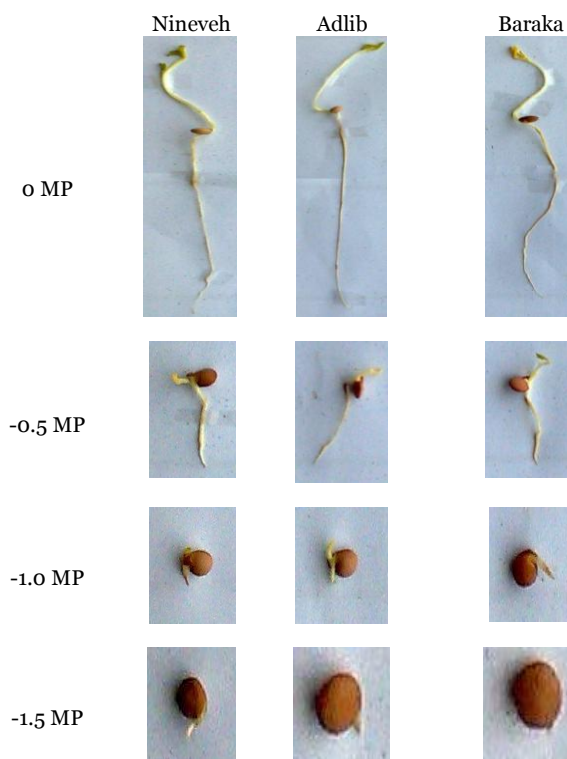


Fig. 3. Nature of germination and seedling performances of three lentil cultivars in response to four osmotic potentials using NaCl concentrations.

The fourth cultivar was Baraka as it showed superiority over Common Vetch in germination energy (35.7%) and (19.9%). Therefore, the worst cultivar was Common Vetch (Fig., 2 and 5). It revealed the lowest values in final germination

percentage (69.7%), germination energy (35%), germination rate (20.8seedling.d⁻¹), radical length (26.6 mm) and plumule length (12.1mm). Cultivar differences in their capabilities of salt tolerance were well established. Unequivocal tolerance discrepancies among cultivars might be attributed to the individual cultivar genome expression ability, techniques that had been applied by producers and pollination contamination of mother plants in the field (Abdel, 2006).

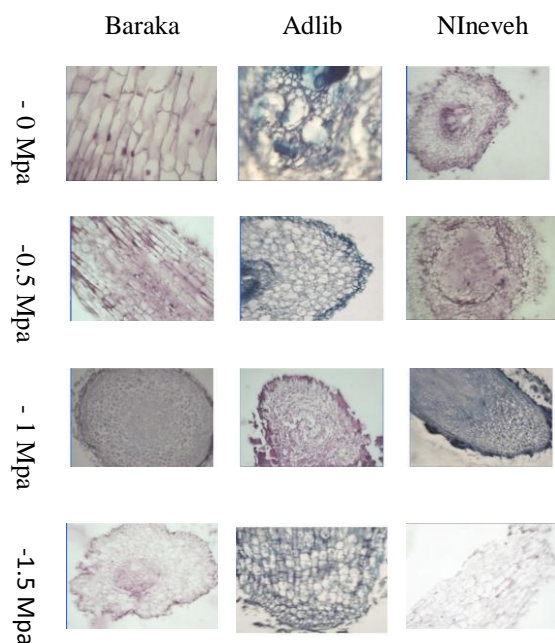


Fig. 4. The influence of varying osmotic levels on root anatomy of three lentil cultivars, Cell destructions are obvious, particularly at higher NaCl rates. (Magnification 7X40).

Varying responses between species were confirmed by Yousif et al. (2010). They examined the difference in the salt tolerance mechanisms between New Zealand spinach and water spinach (*Ipomoea aquatica* L.). Both plants were exposed to salt stress by daily irrigation with 0, 50, 100 and 200 mM NaCl solution for 14 days. They found that the growth of water spinach was markedly and gradually reduced with increasing salinity, whereas that of New Zealand

spinach was increased with elevating salinity, indicating that New Zealand spinach is halophilic.

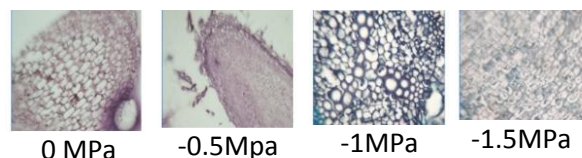


Fig. 5. The influence of varying osmotic levels on root anatomy of Common Vetch local cultivar. Cell destructions are obvious, particularly at higher NaCl rates. (Magnification 7X40).

Cultivar and osmotic solution interactions

Mungbean seeds appeared to be the most potent under all tested osmotic potentials (Table, 1& Figure, 1, 1a, b, c). This cultivar manifested the highest plumule lengths under distilled water (108 mm), -0.5 Mpa (21mm) and -1.5 Mpa (3mm). Moreover this cultivar exhibited the best germination rate: germination percentage ratio and days required for first emergence at all osmotic potentials (Fig. 3). The results also revealed that Adlib cultivar germination performance under distilled water, - 0.5 Mpa and -1Mpa was preponderance over all detected cultivars. It manifested the highest radical lengths (121.25 mm), (21.25mm) and (3 mm), respectively. It is worthy to mention Baraka cultivar overwhelming on all cultivar and all osmotic solutions in germination energy, germination rate, and lowest time for peak germination under 0, -0.5 and -1Mpa. Cultivar differences were obvious at the two highest potentials 0 and -0.5 Mpa. However, as the potential being decreased the variation among cultivar and /or species were gradually vanished. These results suggested that at high potential there were a chance to distinguish cultivars/and or species competitions. On the other hand when salt aggravated, plants lost their salt tolerance capabilities owing to overwhelming salt influences. Exiguously plant responses under low potential might be attributed to

the effects of salts on cell metabolic (Fig. 1, 4), Amini and Ehsanpour (2005) germinated seeds of two tomato cultivars on medium containing only water agar, then transferred to MS medium supplemented with different concentrations of NaCl (0, 40, 80, 120 and 160 mM) for 21 days. They manifested that increasing of salinity resulted in increasing of soluble proteins in stem and leaf of cv. Isfahani but decreasing in cv. Shirazy. Soluble proteins in roots of both cultivars showed some variations.

References

- Abdel CG. 1989.** Drought hardening of onion seeds (*Allium cepa* L. Texas Grano cv.) and its effects on germination capacity under osmotic potential levels. Tec. Res. Apin. 264-74.
- Abdel CG. 2006.** Improving yield and yield quality of four faba bean cultivars grown under rainfalls: 2- Application of growth regulators. Mesopotamia J. of Agric. **34(4)**, 21-30.
- Abdel CG, Salih AW. 1994. Germination Capacity of Fenugreek seeds as influenced by water availabilities. Tech. Res. J. **71(9)**, 71-78.
- Abdel CG, Al-Hamadany SYH. 2007a.** Evaluation of some faba bean (*Vicia faba* L.) cultivars for drought resistance and water consumptive use: 1- Germination of faba bean seeds obtained from water stressed and well irrigated plants. Mesopotamia J. of Agric. **35(1)**, 18-27.
- Abdel CG, Al-Hamadany SYH. 2007b.** Evaluation of some faba bean (*Vicia faba* L.) cultivars for drought resistance and water consumptive use. 2- Effect of supplementary irrigation. Mesopotamia J. of Agric. **35(1)**, 28-36.
- Amini F, Ehsanpour AA. 2005.** Soluble proteins, Proline, Carbohydrates and Na⁺/Cl⁻ changes in two tomato (*Lycopersicon esculentum* Mill.) cultivars under in vitro salt stress. Am. J. Biochem. Biotech. **4**, 212-216.
- Ashraf M. 2001.** Relationships between growth and gas exchange characteristics in some salt-tolerant amphidiploid *Brassica* species in relation to their diploid parents. Environ. Exp. Bot. **45**, 155-163.
- Awad AS, Edwards DG, Campbell LC. 1990.** Phosphorus enhancement of salt tolerance of tomato. Crop Sci. **30**, 123-128.
- Ayala F, O'Leary JW, Schumacher KS. 1996.** Increased vacuolar and plasma membrane H⁺ - ATPase activities in *Salicornia bigelovii* Torr. in response to NaCl stress. Physiologia Plantarum **99**, 328-334.
- Ayers RS, Wescot DW. 1985.** Water quality for Agriculture. Irrigation and Drainage Paper 29, FAO, Rom. p. 174.
- Azooz MM, Ismail AM, Elhamd MFA. 2009.** Growth, lipid peroxidation and antioxidant enzyme activities as a selection criterion for the salt tolerance of Maize cultivars grown under salinity stress. Int. J. Agric. Biol. **11**, 21-26.
- Bassil ES, Kaffka SR. 2002.** Response of safflower to saline soils and irrigation, part I. Consumptive water use. Agric. Water Manage. **54**, 67-80.
- Bayuelo-Jimenez JS, Debouck DG, Lynch JP. 2003.** Growth, gas exchange, water relations, and ion composition of *Phaseolus* species grown under saline conditions. Field Crops Res. **80**, 207-222.
- Ben SL, Alpha MJ, Bahl J, Guillot- Salomon T, Dubacq JP. 1993.** Plant Physiol. Biochem. **31**, 547

557.

Hodson JP, Blumwald E. 2001. Engineering salt-tolerant Brassica plants: Characterization of yield and seed oil quality in transgenic plants with increased vacuolar sodium accumulation. *PNAS* **98(22)**, 12832–12836.

Berlyn GP, Miksche JP. 1976. Botanical Micro Technique and Cytochemistry. Ames Iowa Press.

Borsani O, Valpuesta V, Botella MA. 2001. Evidence for a Role of Salicylic Acid in the Oxidative Damage Generated by NaCl and Osmotic Stress in Arabidopsis Seedlings. *Plant Physiology* **126**, 1024–1030.

Bradford KJ. 1986. Manipulation of seed water relations via osmotic priming to improve germination under stress conditions. *Hort. Sci.* **21**, 1105-1112.

Braun YM, Hassidim HR, Reinhold L. 1986. Studies on H⁺-translocating ATPases in plants of varying resistance to salinity. *Plant Physiology* **81**, 1050-1056.

Schumaker. 2002. Increased vacuolar Na⁺/H⁺ exchange activity in *Salicornia bigelovii* Torr. In response to NaCl. *Journal of Experimental Botany* **53(371)**, 1055-1065.

Brown PH, Shelp BJ. 1997. Boron mobility in plants. *Plant and Soil* **193**, 85–101.

Champagnol F. 1979. Relationships between phosphate nutrition of plants and salt toxicity. *Phosphorus Agri.* **76**, 35-43.

Chen Z, Newman I, Zhou M, Mendham N, Zhang G, Shabala S. 2005. Screening plants for

salt tolerance by measuring K⁺ flux: a case study for barley. *Plant Cell and Environment* **28**, 1230-1246.

Chen ZT, Cuin A, Zhou M, Twomey A, Naidu BP, Shabalam S. 2007. Compatible solute accumulation and stress-mitigating effects in barley genotypes contrasting in their salt tolerance. *Journal of Experimental Botany* **58(15/16)**, 4245–4255.

Craufurd PQ, Wheeler TR, Ellis RH, Summerfield RJ, Williams JH. 1999. Effect of temperature and water deficit on water use efficiency, carbon isotope discrimination, and specific leaf area in peanut. *Crop Sci.* **39**, 136-142.

Cuin TA, Shabala S. 2007. Potassium efflux channels mediate Arabidopsis root responses to reactive oxygen species and the mitigating effect of compatible solutes. *Plant Cell and Environment* **7**, 875–885.

Dat JF, Vandenede F, Vranova E, Montagu MV, Inze D, Breusegem FV. 2000. Dual action of the active oxygen species during plant stress response. *Cells Mol. Life* **57**, 779-95.

Deef HE. 2007. Influence of Salicylic acid on stress tolerance seed germination of *Triticum aestivum* and *Hordeum vulgare*. *Advances Biological Research* **1(1-2)**, 40-48.

Deshpande US, Deshpande SS. 1991. Legumes. In: Salunkhe D.K., Deshpande S.S. (eds): *Foods of Plant*.

Finch-Savage WE, Rowse HR, Dent KC. 2005. Development of combined imbibition and hydrothermal threshold models to simulate maize (*Zea mays*) and chickpea (*Cicer arietinum*) seed germination in variable environments. *New Phytologist* **165**, 825–838.

- Flowers TJ, Yeo AR. 1986.** Ion relations of plant under drought and salinity. *Australian Journal of Plant Physiology* **13**, 75-91.
- Flowers TJ, Troke PF, Yeo AR. 1977.** The mechanisms of salt tolerance in halophytes. *Annual Review of Plant Physiology* **28**, 89-121.
- Frias J, Vidal-Valverde C, Sotomayor C, Diaz-Pollan C, Urbano H. 1999.** Influence of processing on available carbohydrate content and antinutritional factors in chickpeas. *European Food Research and Technology* **210**, 340-345.
- Gama PBS, Inanaga IS, Tanaka K, Nakazawa R. 2007.** Physiological response of common bean (*Phaseolus vulgaris* L.) seedlings to salinity stress. *African Journal of Biotechnology* **6(2)**, 79-88.
- Garg AK, Kim JK, Owens TG, Ranwala AP, Choi YD, Kochian LV, Wu R.J. 2002.** Trehalose accumulation in rice plants confers high tolerance levels to different a biotic stresses. *Proceedings of the National Academy of Sciences USA* **99**, 15898-15903.
- Ghassemi-Golezani K, Aliloo AA, Valizadeh M, Moghaddam M. 2008.** Effects of hydro and osmo-priming on seed germination and field emergence of lentil (*Lens culinaris* Medik.). *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* **36(1)**, 29-33.
- Glenn E, Brown JJ, Blumwald E. 1999.** Salt tolerance and crop potential of halophytes. *Crit Rev. Plant Sci.* **18**, 227-256.
- Graham EP, Dietrich MA, Schumaker KS. 2002.** Increased vacuolar Na⁺/H⁺ exchange activity in *Salicornia bigelovii* Torr. In response to NaCl. *Journal of Experimental Botany*, 53(371): 1055-1065.
- Grattan SR, Grieve CM. 1999.** Salinity-mineral nutrient relations in horticultural crops. *Sci. Hort.* **78**, 127-157.
- Górecki RJ, Piotrowicz-Cieslak AI, Obendorf RL. 1997a.** Soluble sugars and flatulence-producing oligosaccharides in maturing yellow lupine (*Lupinus luteus* L.) seeds. *Seed Science Research* **7**, 185-193.
- Greenway H, Osmond CB. 1972.** Salt responses of enzymes from species differing in salt tolerance. *Plant Physiology* **49**, 256-259.
- Greenway H, Munns R. 1983.** Interactions between growth, uptake of Cl and Na and water relations of plants in saline environments. *Plant Cell Environ.* **6**, 575-589.
- Harris D. 2001.** On-farm seed priming: A key technology to improve the livelihoods of esource-poor farmers in marginal environments. Bangor: UK Department for International Development and University of Wales.
- He L, Gao Z, Li R. 2009.** Pretreatment of seed with H₂O₂ enhances drought tolerance of wheat (*Triticum aestivum* L.) seedlings *African Journal of Biotechnology* **8(22)**, 6151-6157.
- Heydecker W, Coolbaer P. 1977.** Seed treatments for improved performance survey and attempted prognosis. *Seed Sci. Technol.* **5**, 353-425.
- Horie T, Schroeder IJ. 2004.** Sodium Transporters in Plants. *Diverse Genes and Physiological Functions.* *Plant Physiology.* American Society of Plant Biologists **136**, 2457-2462.
- Horie T, Schroeder IJ. 2004.** Sodium Transporters in Plants. *Diverse Genes and*

Physiological Functions. *Plant Physiology* **136**, 2457–2462.

Hung SH, Yu CW, Lin CH. 2005. Hydrogen peroxide functions as a stress signal in plants. *Bot. Bull. Acad. Sin.* **46**, 1-10.

Inge SM, Gilliam M, Jha D, Mayo GM, Roy SJ, Coates JC, Haseloff J, Testerb M. 2009. Shoot Na⁺ Exclusion and Increased Salinity Tolerance Engineered by Cell Type Specific Alteration of Na⁺ Transport in Arabidopsis. *The Plant Cell Preview*, American Society of Plant Biologists.

Iyengar ERR, Reddy MP. 1996. Photosynthesis in highly salt-tolerant plants. In: M. Pessaraki (ed.), *Handbook of Photosynthesis*, Marcel Dekker, New York, p. 897-909.

Johnson SE, Lauren JG, M. Welch R, Duxbury JM. 2005. A comparison of the effects of micronutrient seed priming and soil fertilization on the mineral nutrition of chickpea (*Cicer arietinum*), lentil (*Lens culinaris*), rice (*Oryza sativa*) and wheat (*Triticum aestivum*) in Nepal. *Expl Agric.* **41**, 427–448.

Kadlec P, Doslov J, Ško JB, Skulino M. 2003. Degradation of α -Galactosides during the germination of Grain Legume Seeds. *Czech J. Food Sci.* **26(2)**, 99–108.

Kadlec P, Dostalova J, Bernaskova J, Skulinova M. 2008. Degradation of α -Galactosides during the Germination of Grain Legume Seeds. *Czech J. Food Sci.* **26(2)**, 99–108.

Kafkafi U, Siddiqi MY, Ritchie RJ, Glass ADM, Ruth TJ. 1992. Reduction of nitrate (¹³NO₃) influx and nitrogen (¹³N) translocation by tomato and

melon varieties after short exposure to calcium and potassium chloride salts. *J. Plant Nutr.* **15**, 959-975.

Katiyar-Agarwal SJ, K. Kim MA, Fu X, Huang A, Zhu JK. 2006. The plasma membrane Na⁺/H⁺ antiporter SOS1 interacts with RCD1 and functions in oxidative stress tolerance in Arabidopsis. *Proc Natl Acad Sci. USA* **103**, 18816-18821.

Kaya MD, Okcu G, Atak M, Cıkkılı Y, Kolsarici O. 2006. Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus annuus* L.). *Eur. J. Agron.* **24**, 291-295.

Kaydan D, Yagmur M. 2008. Germination, seedling growth and relative water content of shoot in different seed sizes of triticale under osmotic stress of water and NaCl. *African Journal of Biotechnology* **7(16)**, 2862-2868.

Kishor PBK, Hong Z, Miao GH, Hu CAA, Verma DPS. 1995. Over-expression of Δ 1-pyrroline-5-carboxylate synthetase increases proline production and confers osmotolerance in transgenic plants. *Plant Physiol.* **108**, 1387–1394.

Koster KL, Leopold AC. 1988. Sugars and desiccation tolerance in seeds. *Plant Physiology* **88**, 829–832.

Liu J, Zhu JK. 1998. A calcium sensor homolog required for plant salt tolerance. *Science* **280**, 1943-1945.

Lucas WJ, Alexande JM. 1981. Influence of turgor pressure manipulation on plasmalemma transport of HCO₃⁻ and OH⁻ in *Chara corallina*. *Plant Physiology* **68**, 553-559.

Marschner H. 1995. Mineral Nutrition of Higher Plants. 2nd ed. Academic Press, London, p. 889.

Materne M, McNeil D, Hobson K, Ford R. 2007. Abiotic stresses of lentils, In Yadav S.S., D. McNeil and P. C. Stevenson eds, Lentil: An Ancient Crop for Modern Times. Springer, Dordrecht, p. 315-330.

Mohamed AN, Rahman MH, Alsadon AA, Islam R. 2007. Accumulation of Proline in NaCl-treated Callus of Six Tomato (*Lycopersicon esculentum* Mill.) Cultivars. Plant Tissue Cult. & Biotech. **17(2)**, 217-220.

Muehlbauer FJ, Cubero JI, Summerfield RJ. 1985. Grain Legume Crops. p. 266-311. In: Lentil (*Lens culinaris* Medic.). Summerfield R. J. and E. H. Roberts (Eds.). Collins, 8 Grafton Street, London, UK.

Munns R. 1993. Physiological processes limiting plant growth in saline soil: some dogmas and hypotheses. Plant Cell and Environment **16**, 15-24.

Munns R, Tester M. 2008. Mechanisms of salinity tolerance. Annu. Rev. Plant Biol. **59**, 651-681.

Munns R, Passioura JB. 1984. Effect of prolonged exposure to NaCl on the osmotic pressure of leaf xylem sap from intact, transpiring barley plants. Australian Journal of Plant Physiology **11**, 497-507.

Murungu FS, Chiduza C, Nyamugafata P, Clark LJ, Whalley WR, Finch-Savage WE. 2004. Effects of sowing occasion and 'on-farm seed priming' on emergence and growth of maize in semi-arid Zimbabwe. Field Crops Research **89**, 49-59.

Musa AM, Harris D, Johansen C, Kumar J. 2001. Short duration chickpea to replace fallow after aman rice: the role of on-farm seed priming in the

High Barind Tract of Bangladesh. Experimental Agriculture **37**, 509-521.

Noctor G, Foyer CH. 1998. Ascorbate and glutathione: keeping active oxygen under control. Annual Review of Plant Physiology and Plant Molecular Biology **49**, 249-279.

O'Leary JW. 1995. Adaptive components of salt tolerance. In: Handbook of plant and crop physiology 577-588.

Okçu G, Kaya MD, Atak M. 2005. Effects of salt and drought stresses on germination and seedling growth of pea (*Pisum sativum* L.). Turk. J. Agric. For. **29**, 237-242.

Omami EN. 2005. Response of Amaranth to Salinity Stress. (Chapter 4 Differences In Salinity Stress Tolerance In Terms of Growth and Water Use Efficiency Among Four Amaranth Genotypes). Univ. of Pretoria etd. p. 86-115.

Pang XP, Letey J. 1998. Development and evaluation of ENVIRO-GRO, an integrated water, salinity, nitrogen model. Soil Sci. Soc. Am. J. **62**, 1418-1427.

Parida AK, Das AB, Mitra B. 2004. Effects of salt on growth, ion accumulation, photosynthesis and leaf anatomy of the mangrove, *Bruguiera paeviflora*. Trees **18**, 167-174.

Peárez-Alfocea F, Balibrea ME, Santa A, Estan MT. 1996. Agronomical and physiological characterization of salinity tolerance in a commercial tomato hybrid. Plant Soil **180**, 251-257.

Pérez-Alfocea F, Estañ MT, Caro M, Guerrier G. 1993. Osmotic adjustment in *Lycopersicon esculentum* and *L. pennellii* under NaCl and

polyethylene glycol 6000 iso-osmotic stresses. *Physiologia Plantarum* **87**, 493-498.

Pridham JB, Dey PM. 1974. The nature and function of higher plant α -galactosidases. In: *Plant Carbohydrate Biochemistry*. Academic Press, New York: p. 83-96.

Rai SP, Luthra R, Kumar S. 2003. Salt-tolerant mutants in glycolytic salinity response (GSR) genes in *Catharanthus roseus*. *Theor. Appl. Genet.* **106**, 221-230.

Ransome LS, Dowdy RH. 1987. Soybean growth and boron distribution in a sandy soil amended with scrubber sludge. *Journal of Environmental Quality* **16**, 171-175.

Ruan S, Xue D, Tylkowski K. 2000. The influence of priming on germination of rice (*Oryza sativa* L.) seed and seedling emergence and performance in flooded soil. *Seed sci. Tech.* **30**, 61-67.

Sadeghian SY, Yavari N. 2004. Effect of water-deficit stress on germination and early seedling growth in sugar beet. *J. Agron. Crop Sci.* **190**, 138-144.

Saglam S, Day S, Kaya G. 2010. Hydropriming Increases Germination of Lentil (*Lens culinaris* Medik.) under Water Stress. *Not Sci. Biol.* **2(2)**, 103-106.

Santos V, Calil AC, Ruiz HA, Alvarenga EM, Santos CM. 1992. Efeito do estresse salino e hídrico na germinação e vigor das sementes de soja. *Revista Brasileira de Sementes* **14**, 189-194.

Saxena NP, Saxena MC, Ruckenbauer P, Rana RS, El-Fouly M, Shabana R. 1994. Screening techniques and sources of tolerance to salinity and

mineral nutrient imbalances in cool season food legumes. *Euphytica* **73**, 85-93.

Scorer KN, Epel BL, Waisel Y. 1985. Interactions between Mild NaCl Stress and Red Light during Lettuce (*Lactuca sativa* L. cv Grand Rapids) Seed Germination. *Plant Physiol.* **79**, 149-152.

Serrano R, Rodriguez-Navarro A. 2001. Ion homeostasis during salt stress in plants. *Current Opinion in Cell Biology* **13**, 399-404.

Shabala S, Shabala L, van Volkenburgh E. 2003. Effect of calcium on root development and root ion fluxes in salinized barley seedlings. *Functional Plant Biology* **30**, 507-514.

Shalata A, Neumann PM. 2001. Exogenous ascorbic acid (Vitamin C) increases resistance to salt tolerance and reduced lipid peroxidation. *J. Exp. Bot.* **364**, 2207-2211.

Shalhevet J, Hsiao C. 1986. Salinity and drought. A comparison of their effects on osmotic adjustment, assimilation, transpiration and growth. *Irrig. Sci.* **7**, 249-264.

Sharma SN, Prasad R. 1984. Effect of soil moisture regimes on the yield and water use of lentil (*Lens culinaris* Medik). *Irrig. Sci.* **5**, 285-293.

Sharpley AN, eisinger JJ, Power JF, Suarez DL. 1992. Root extraction of nutrients associated with long-term soil management. In: Stewart, B. (Ed.), *Advances in Soil Science*, vol. 19. Springer, p. 151-217.

Shaviv A, Hazan O, Neumann PM, Hagin J. 1990. Increasing salt tolerance of wheat by mixed ammonium nitrate nutrition. *J. Plant Nutr.* **13**, 1227-1239.

Shen B, Jensen. G, Bohnert HJ. 1997. Mannitol protects against oxidation by hydroxyl radicles. *Plant Physiol.* **115**, 527-532.

Silberbush M, Lips SH. 1991. Potassium, nitrogen, ammonium/nitrate ratio, and sodium chloride effects on wheat growth. I. Shoot and root growth and mineral composition. *J. Plant Nutr.* **14**, 751-764.

Sohan D, Jasoni R, Zajicek J. 1999. Plant-water relations of NaCl and calcium-treated sunflower plants. *Environ. Exp. Bot.* **42**, 105-111.

Sung JM, Chiu KY. 1995. Hydration effects on seedling emergence strength of watermelon seed differing in ploidy. *Plant Sci.* **110**, 21-26.

Thumma BR, Naidu BP, Chandra A, Cameron DF, Bahnisch LM, Liu C. 2001. Identification of causal relationship among traits related to drought resistance in *Stylosanthes scabra* using QTL analysis. *J. Exp. Bot.* **52**, 203-214.

Townend J, Mtakwa PW, Mullins CE, Simmonds LP. 1996. Soil physical factors limiting establishment of sorghum and cowpea in two contrasting soil types in the semi-arid tropics. *Soil and Tillage Research* **40**, 89-106.

Wu J, Seliskar DM, Gallagher JL. 1998. *Physiol. Plant* **102**, 307-317. Ben Rai S. L., M. J. Alpha, J. Bahl, T. Guillot-Salomon and J. P. Dubacq (1993). *Plant Physiol. Biochem.* **31**, 547-557.

Yamaguchi TS, Apse MP, Shi H, Blumwald E. 2003. Topological analysis of a plant vacuolar Na⁺/H⁺ antiporter reveals a luminal C terminus that regulates antiporter cation selectivity. *PNAS* **100(21)**, 12510-12515.

Yousif BS, Nguyen NT, Fukuda Y, Hakata H, Okamoto Y, Masaoka Y, Saneokai H. 2010. Effect of Salinity on Growth, Mineral Composition, Photosynthesis and Water Relations of Two Vegetable Crops; New Zealand Spinach (*Tetragonia tetragonioides*) and Water Spinach (*Ipomoea aquatica*). *Int. J. Agric. Biol.* **12(2)**.

Zhang HX, Hodson JN, Williams JP, Blumwald E. 2001b. Engineering salt-tolerant Brassica plants: Characterization of yield and seed oil quality in transgenic plants with increased vacuolar sodium accumulation, *PNAS* **98(22)**, 12832-12836

Zhu JK. 2001. Plant Salt Tolerance. *Trends Plant Sci.* **6**, 66-71.